



Figure 2: 2'-O-Me substituted Amberzyme Enzymatic Nucleic Acid Motif

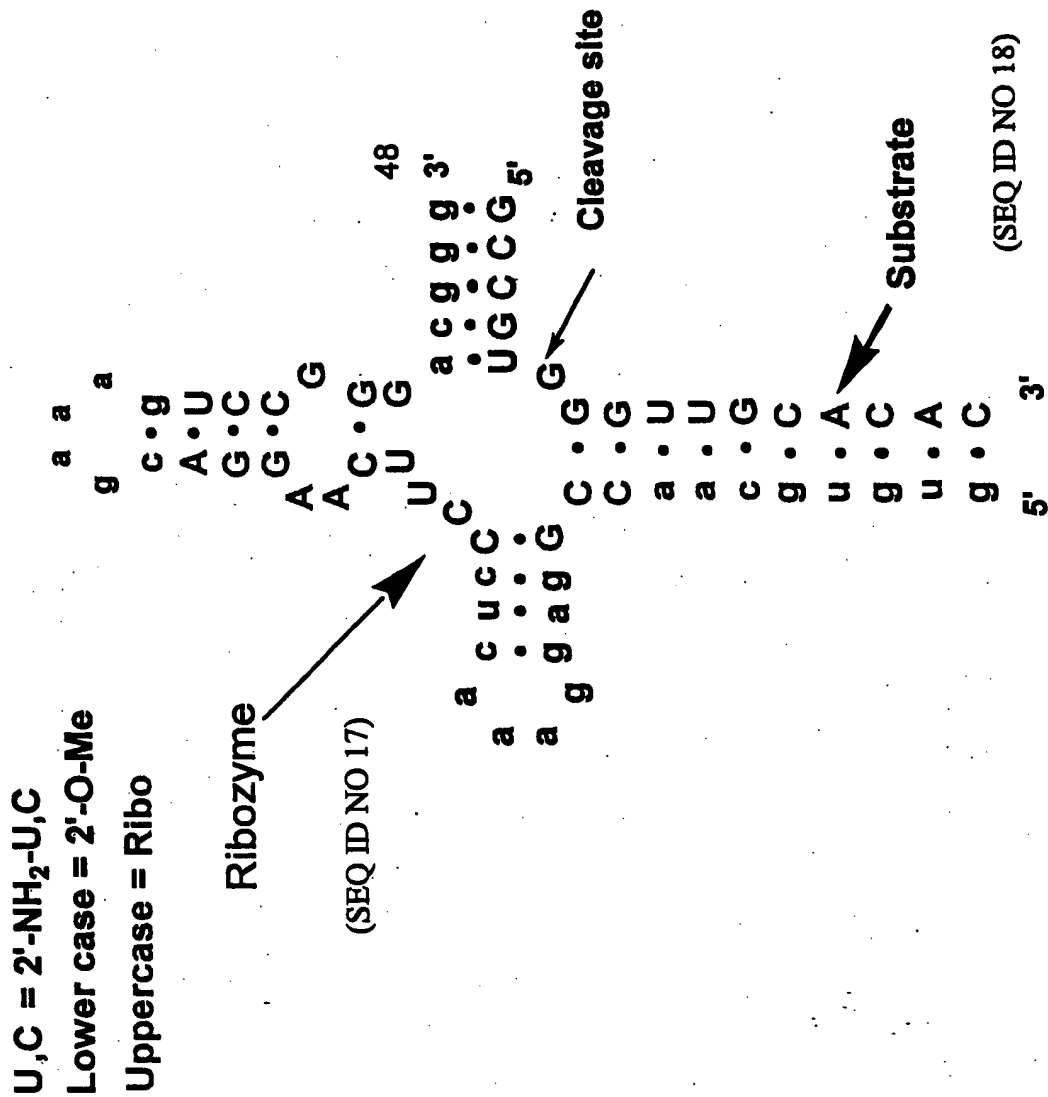
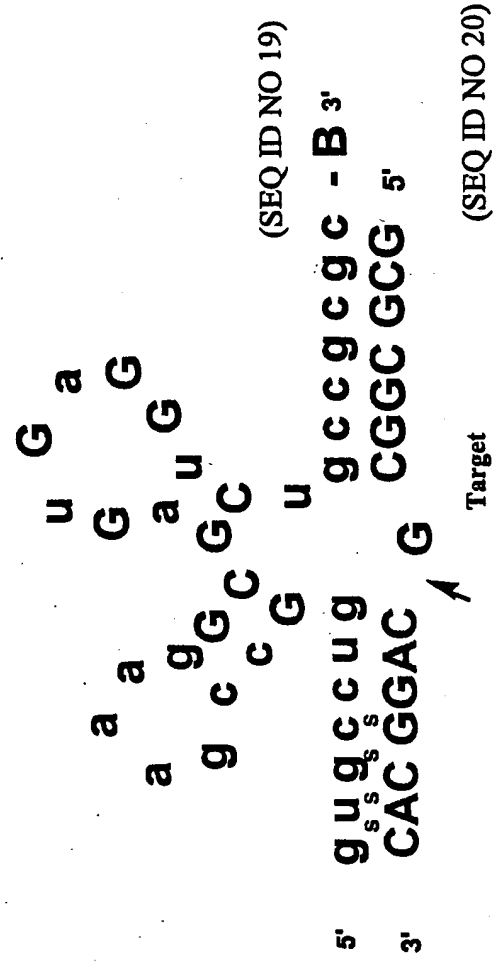


Figure 3: Stabilized Zinzyme Ribozyme Motif

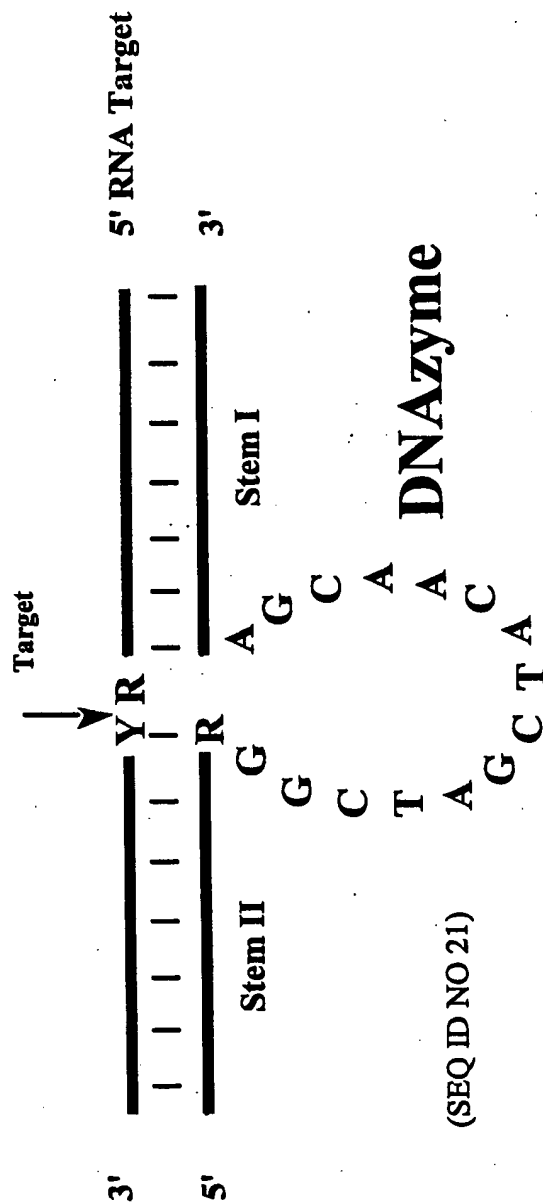
Zinzyme A-motif RZ



Legend

- Uppercase indicates natural ribo residues
- C** indicates 2' - d-NH<sub>2</sub>-C
- Lowercase: 2'-O- Me
- Subscript **s** indicates phosphothioate linkage
- B: 3'-3' abasic moiety

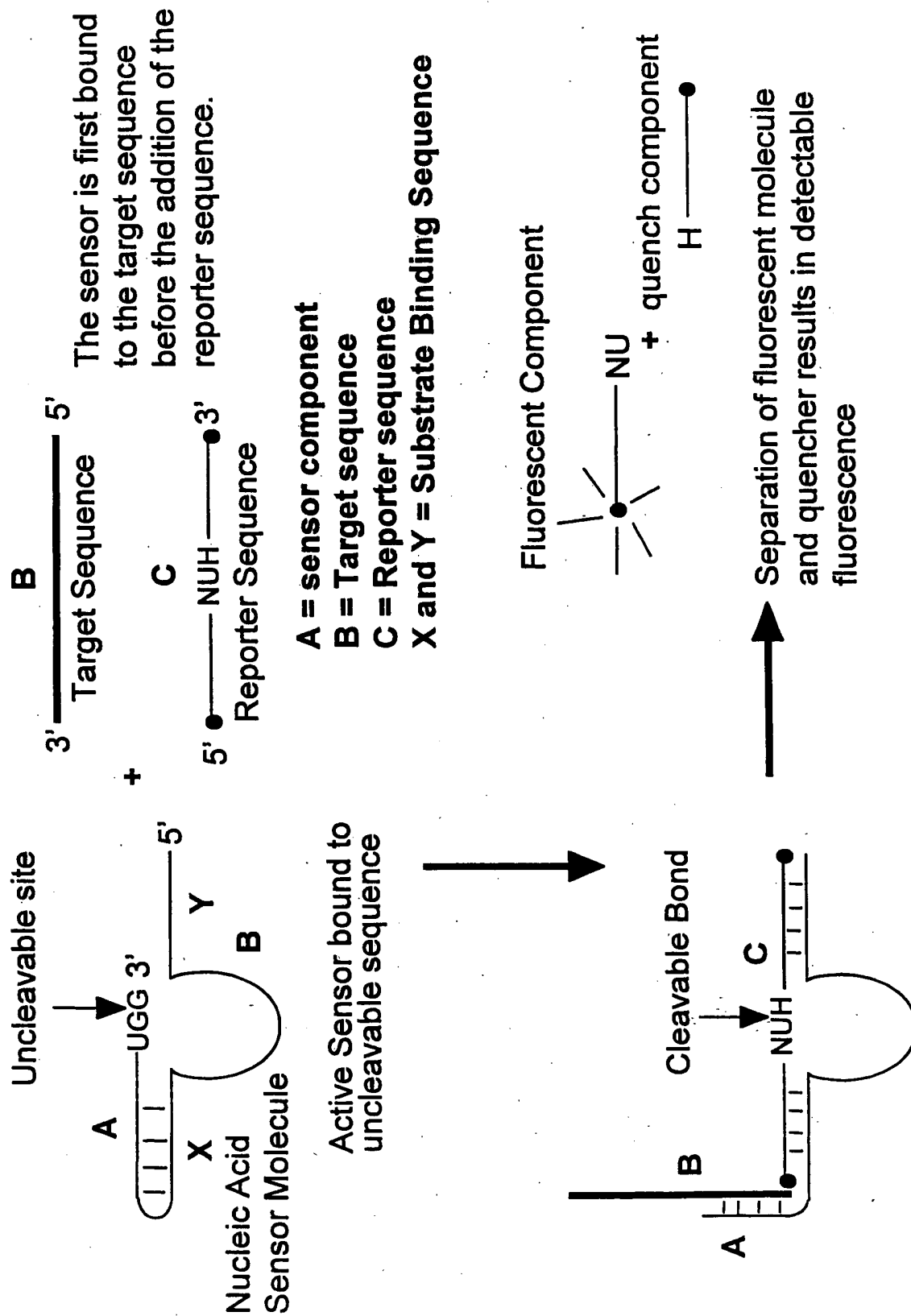
Figure 4: DNase Motif



Legend

Y = U or C  
R = A or G

**Figure 5. Detection of Target Sequence Using a Cis-Blocking Sequence**



**Figure 6. Schematic Diagram Representing the Two Primary Configurations of the Diagnostic effector molecule**

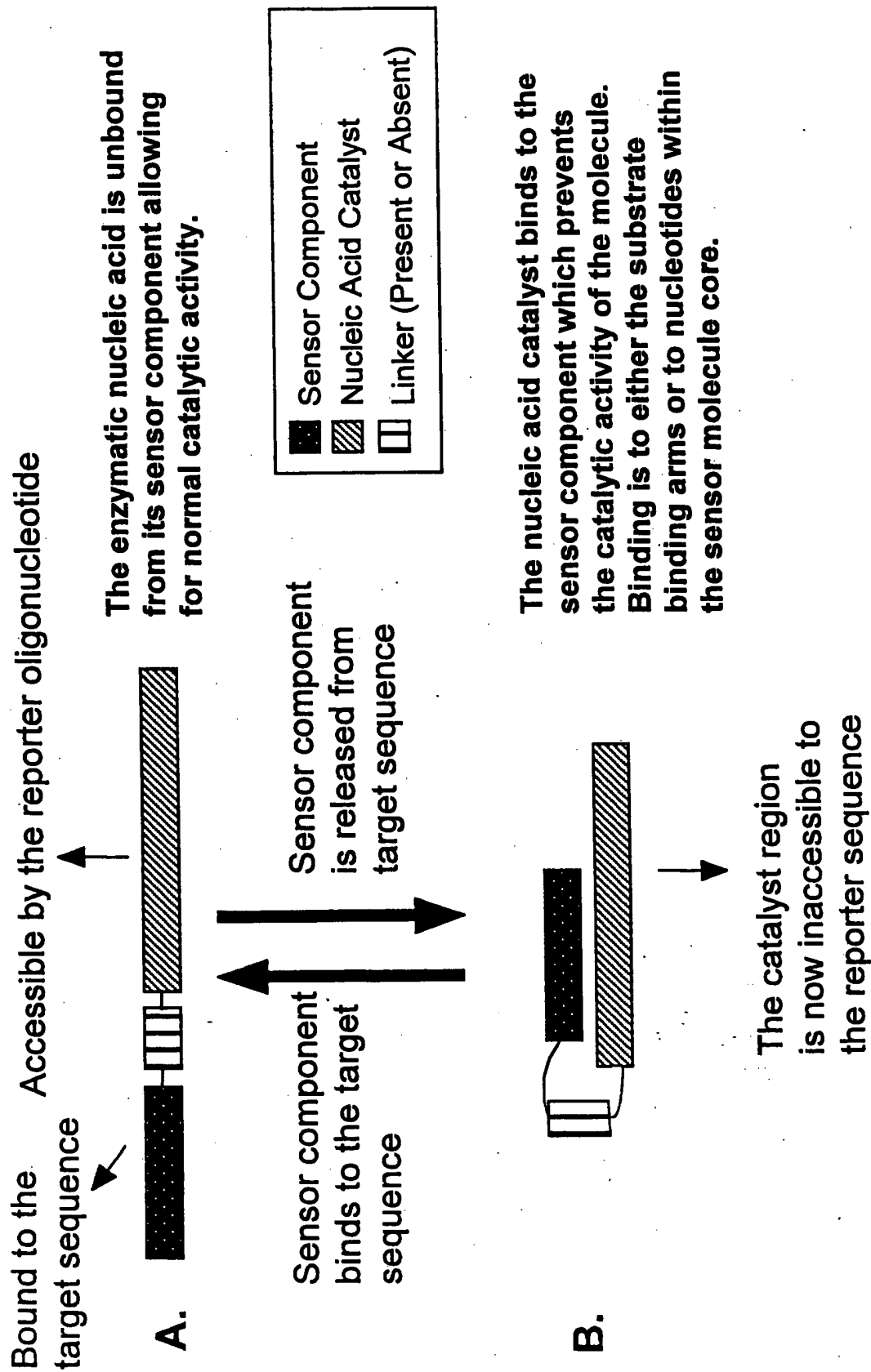


Figure 7. Examples of Nucleic Acid Sensor Molecules

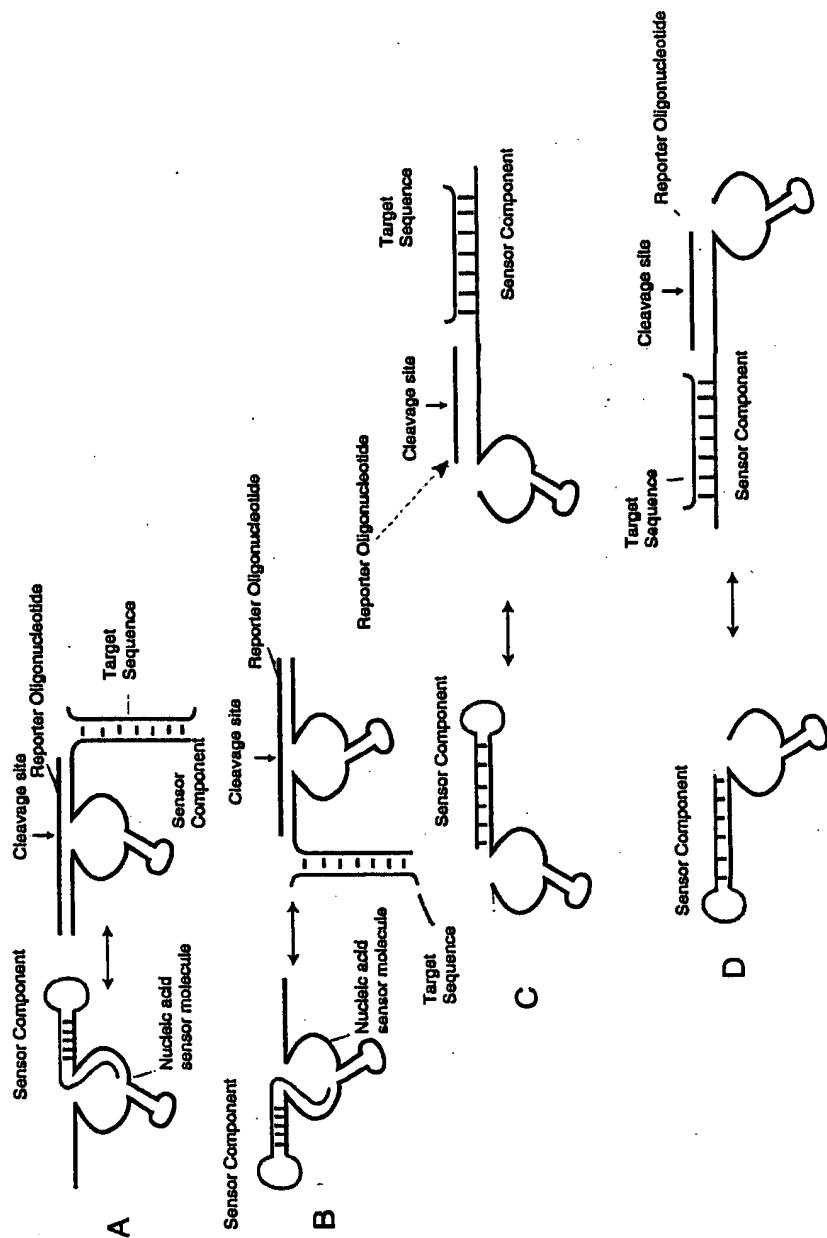


Figure 8. Examples of Nucleic Acid Sensor Molecules

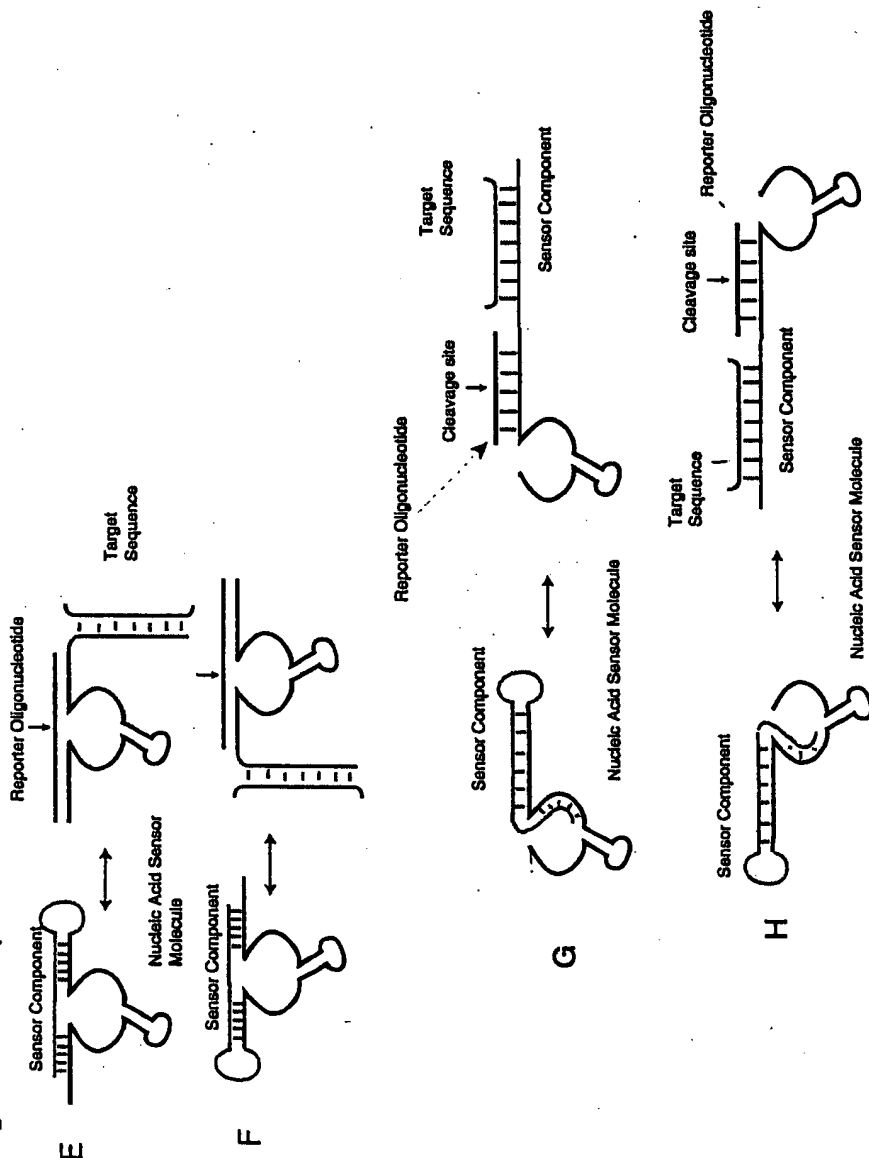




Figure 9. Examples of Nucleic Acid Sensor Molecules

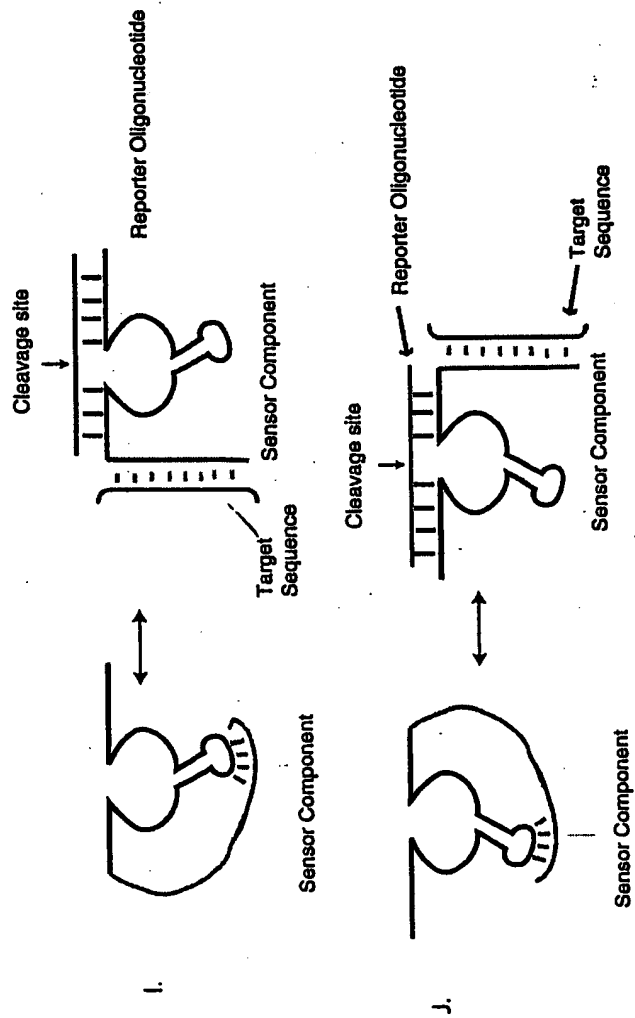
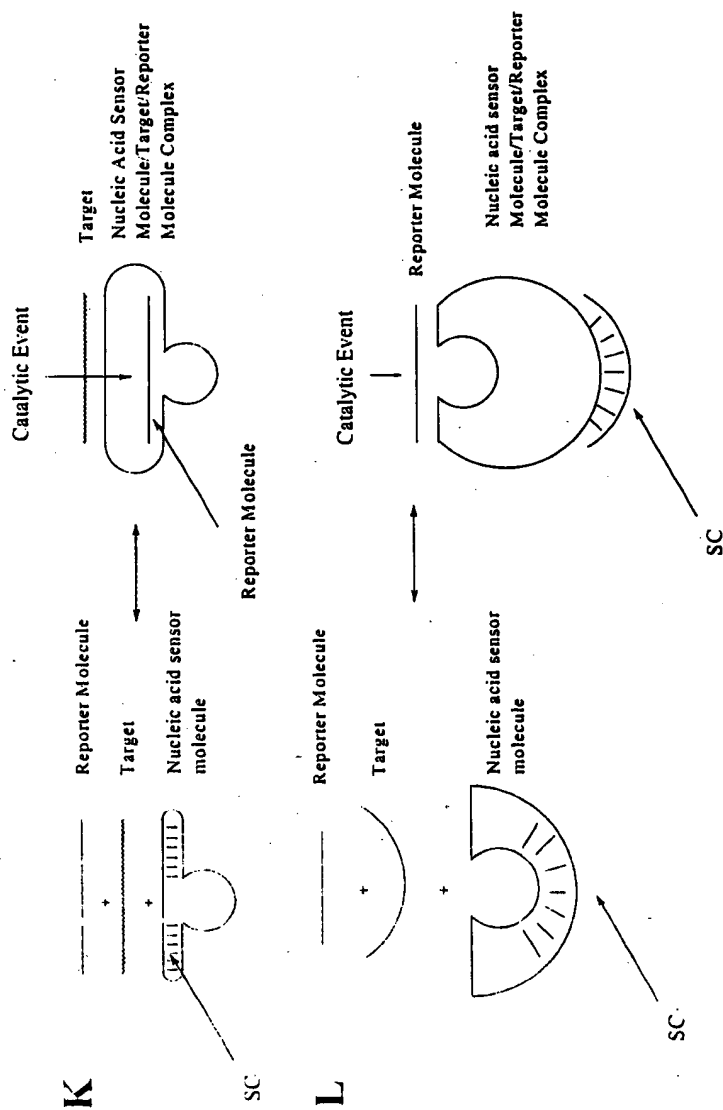
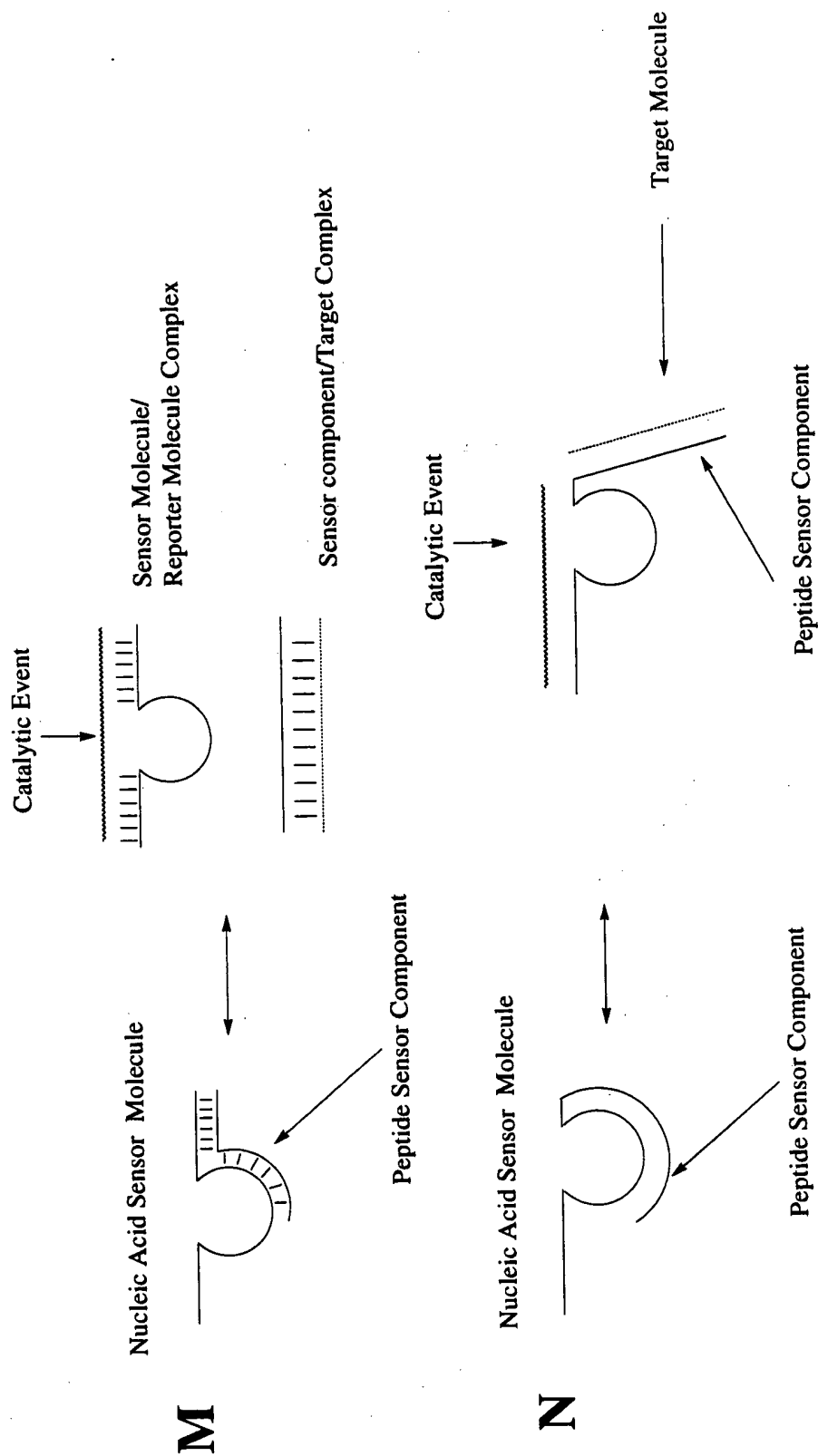


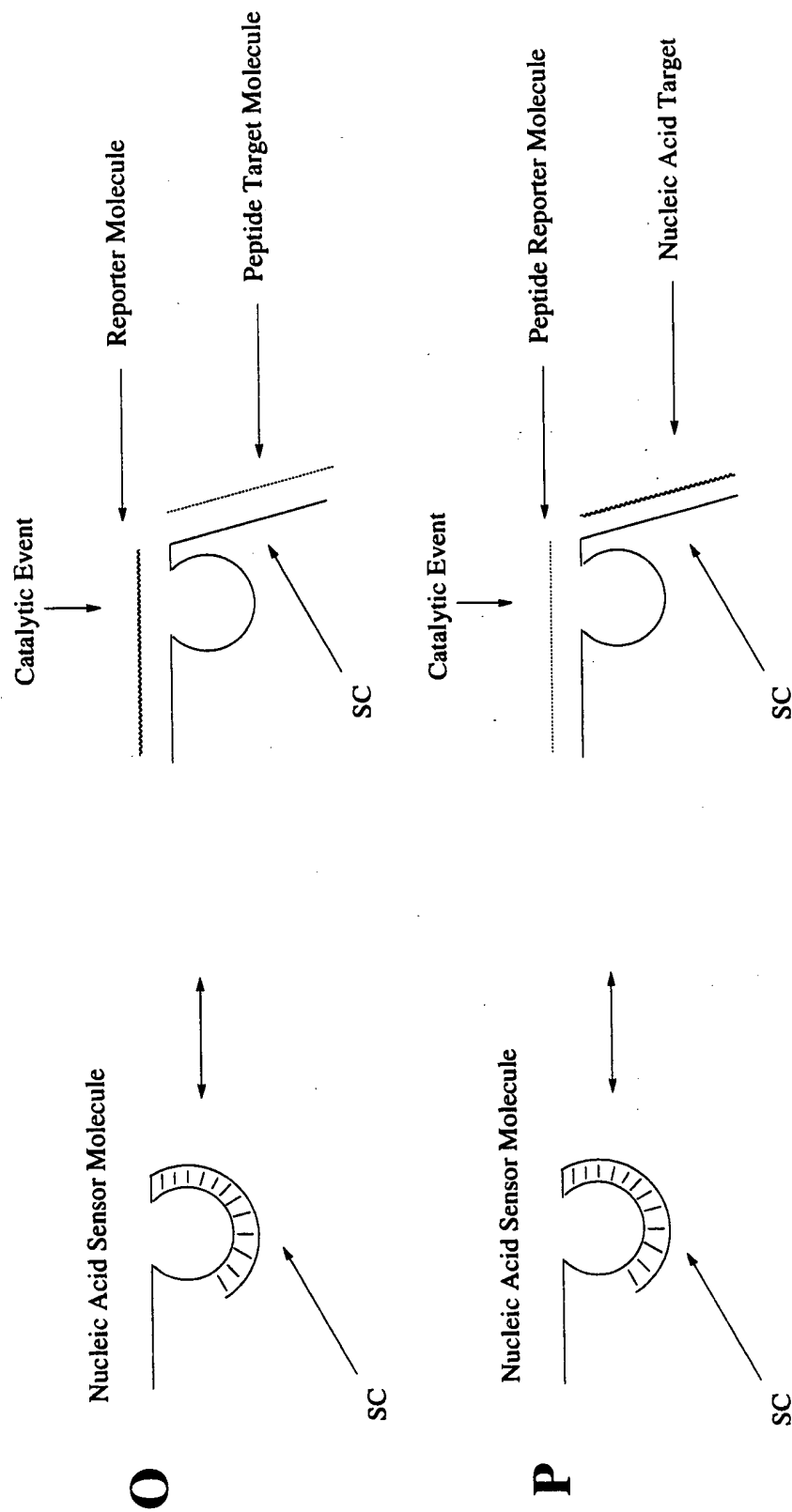
Figure 10: Examples of Nucleic Acid Sensor Molecules



**Figure 11: Examples of Nucleic Acid Sensor Molecules**

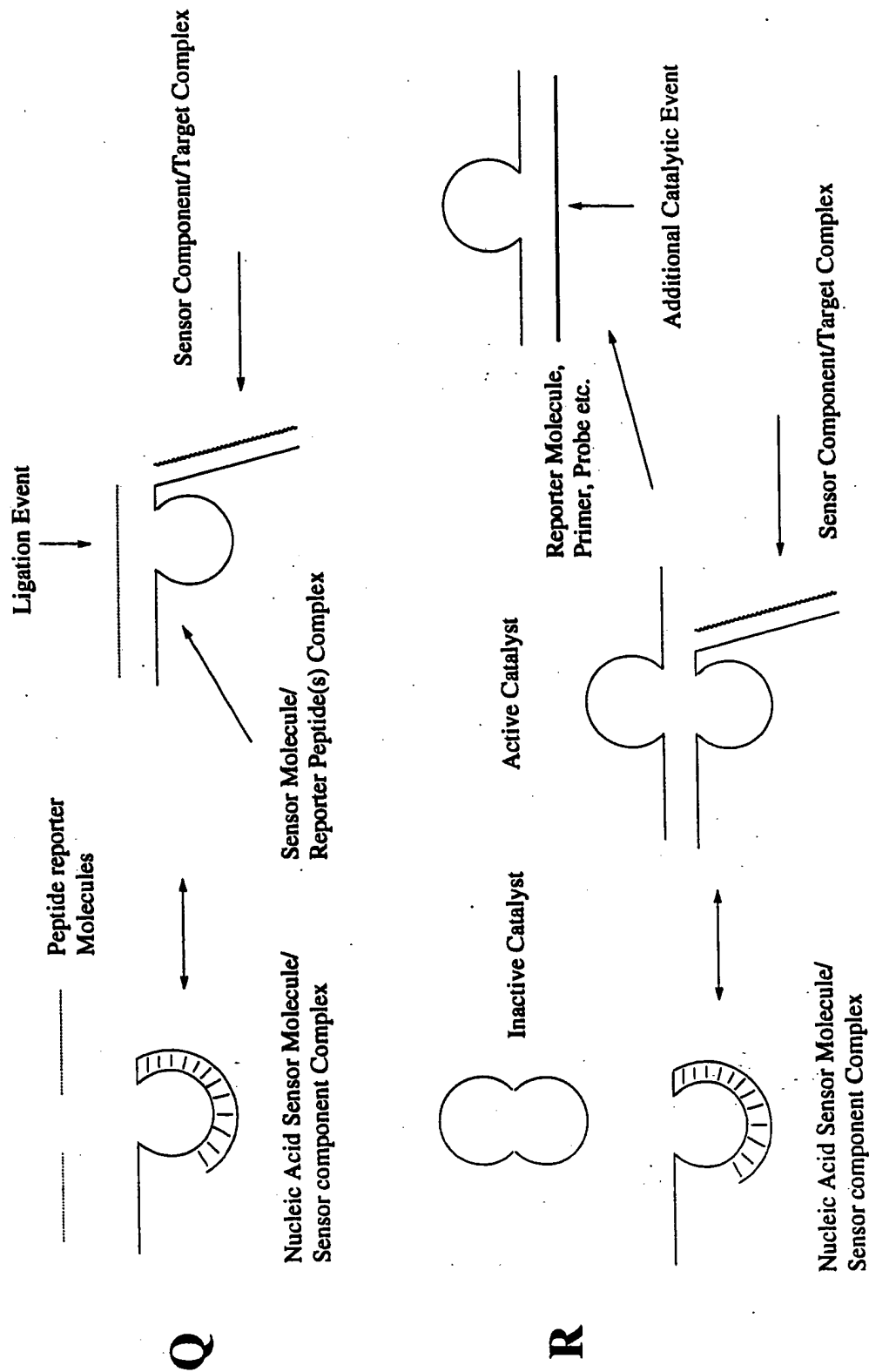


**Figure 12: Examples of Nucleic Acid Sensor Molecules**

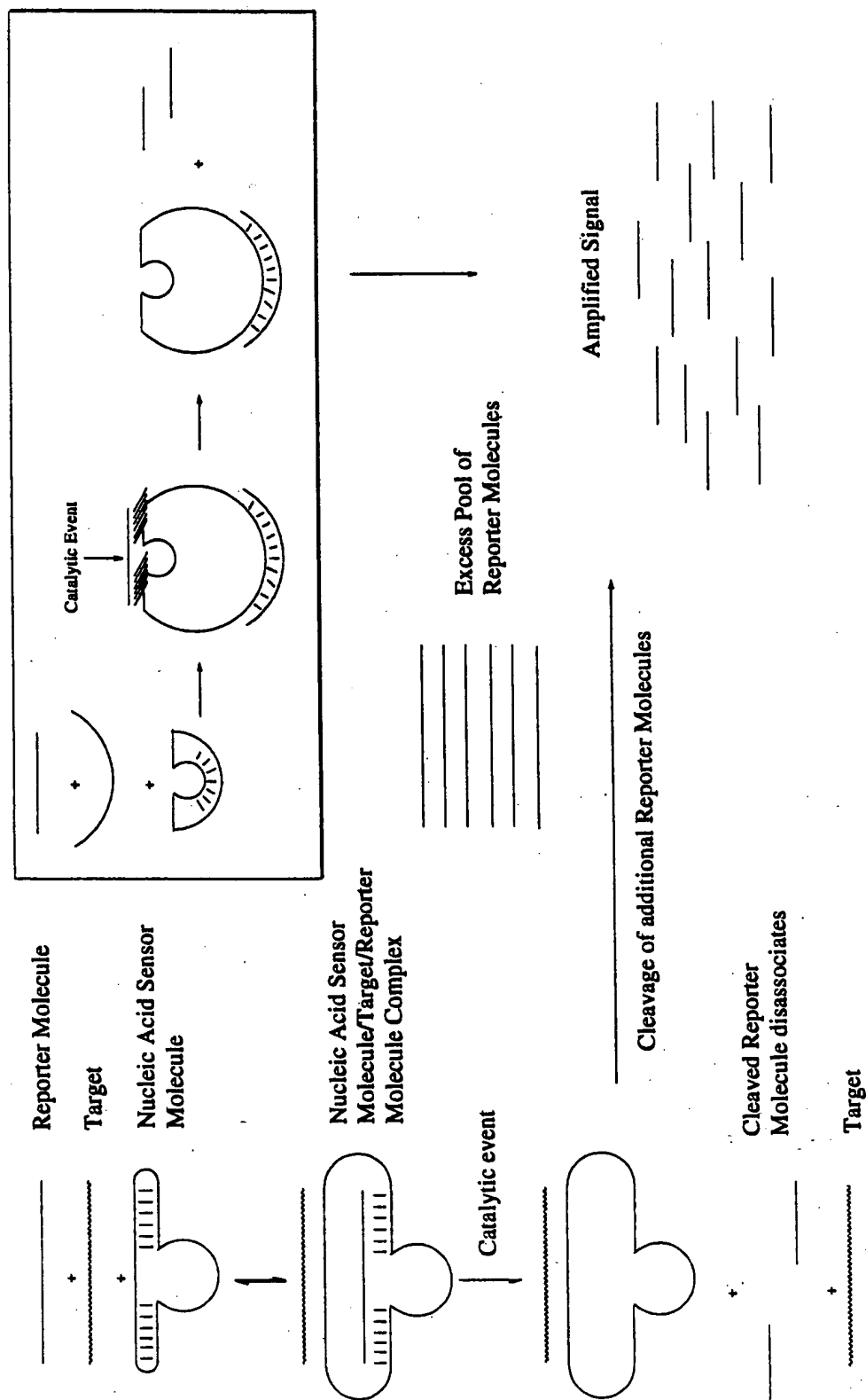


SC = Sensor Component

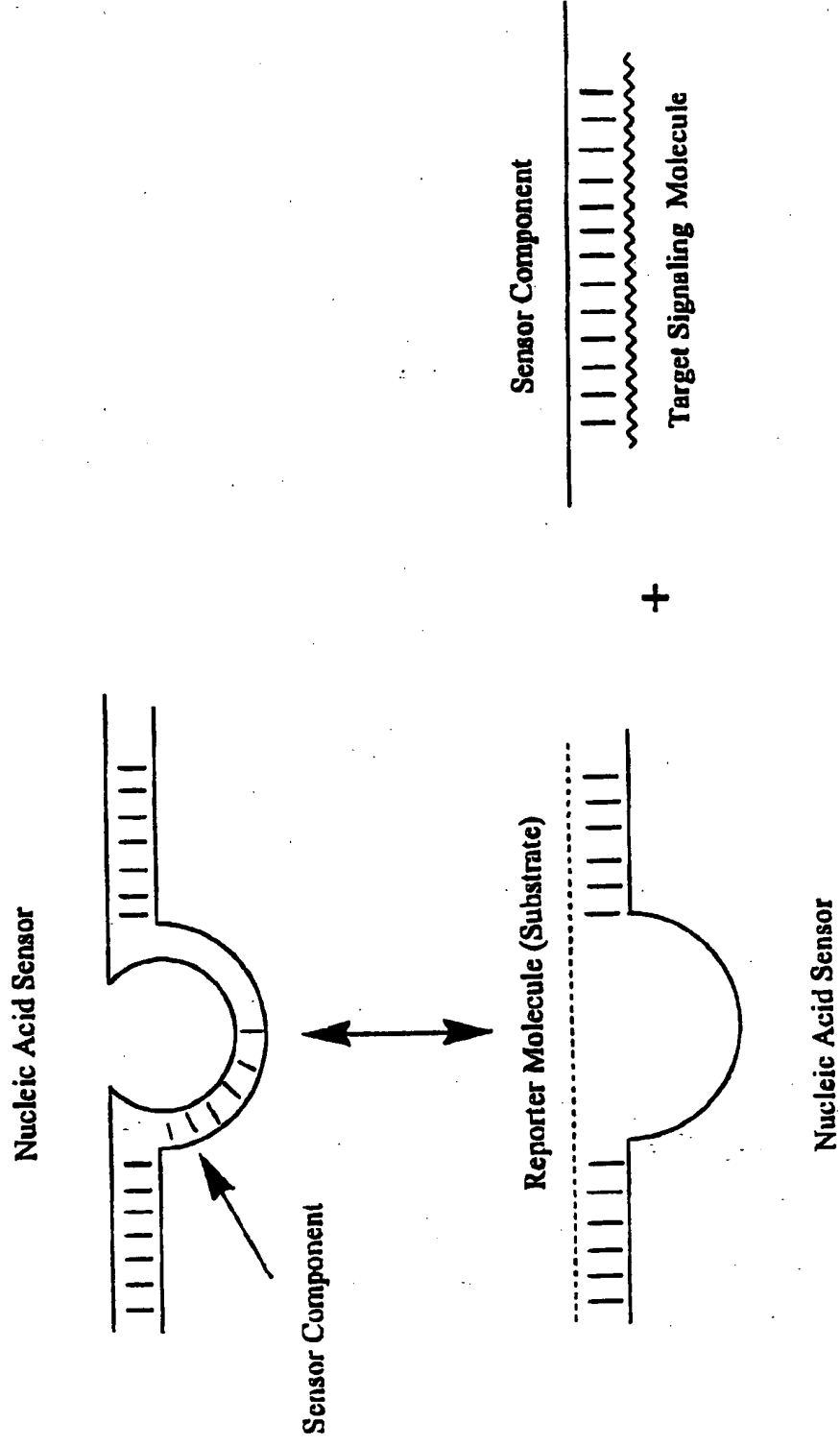
**Figure 13: Examples of Nucleic Acid Sensor Molecules**



**Figure 14: Inherent Amplification of Signal**

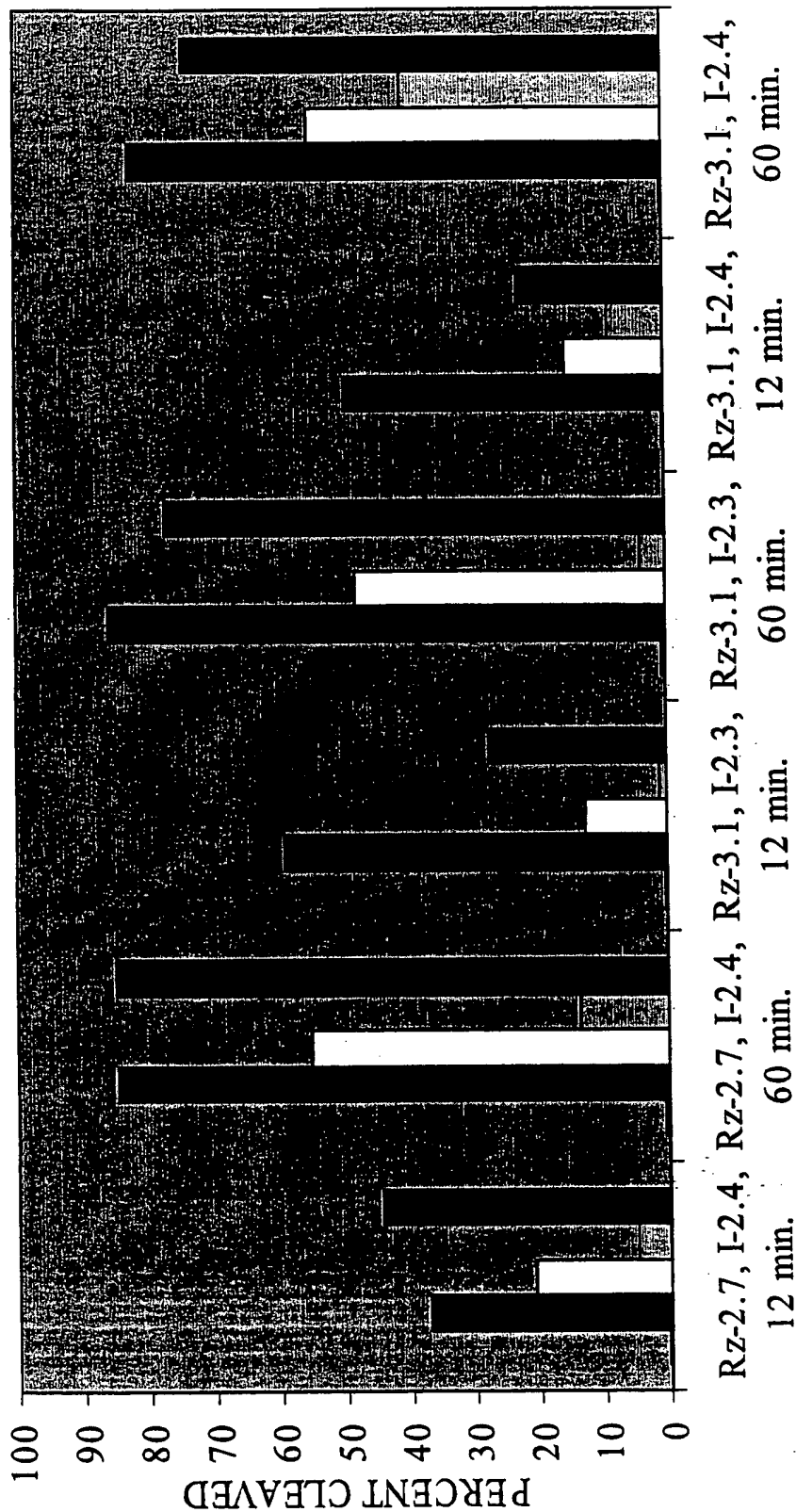


**Figure 15: Example of Diagnostic System**



**Figure 16: Ribozyme Diagnostic Screen**

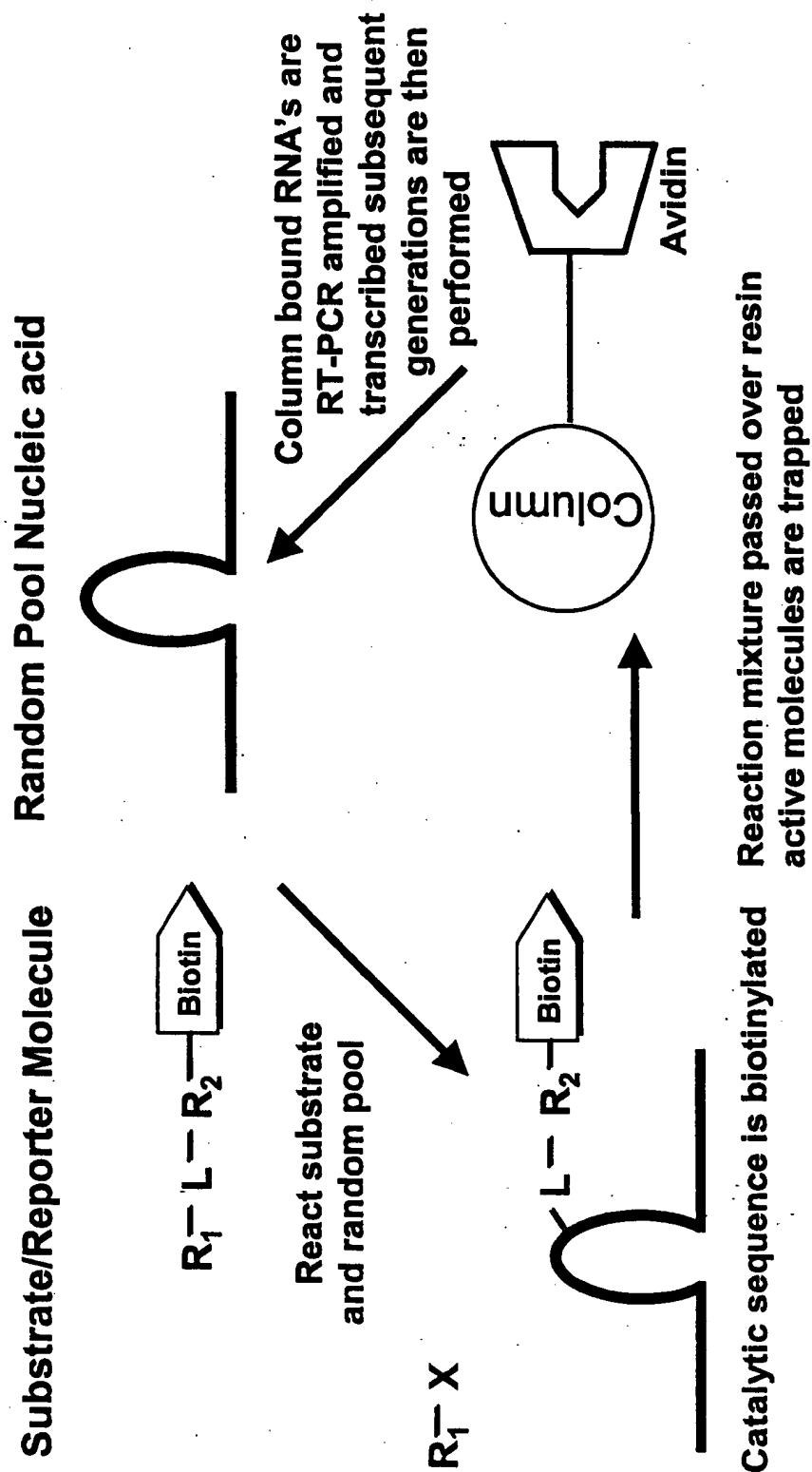
INHIBITORY FOLDING WITH TARGET RESCUE



No Rz 
  +Rz @ 10 nM 
  +Rz, +I @ 20 nM 
  +Rz, +I, +T @ 500 nM



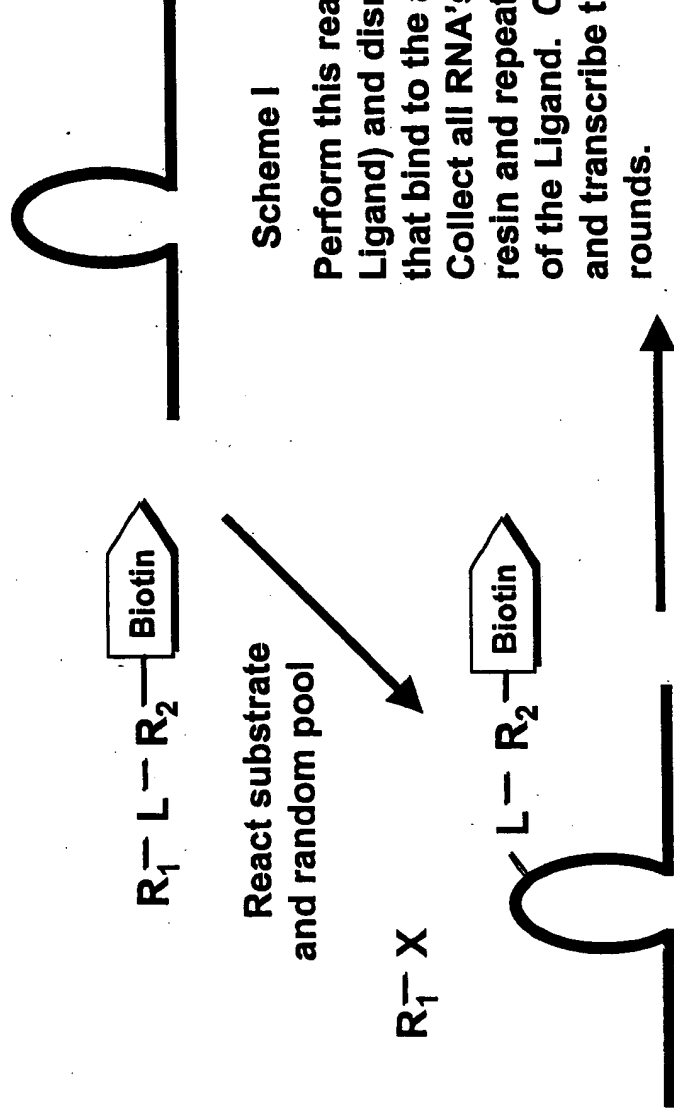
**Figure 17a: Auto-ligation Nucleic Acid Sensor Molecules - Selection Scheme**



## Figure 17b: Auto-ligation Nucleic Acid Sensor Molecules - Ligand Dependent

Substrate/Reporter Molecule + Random Pool Nucleic acid

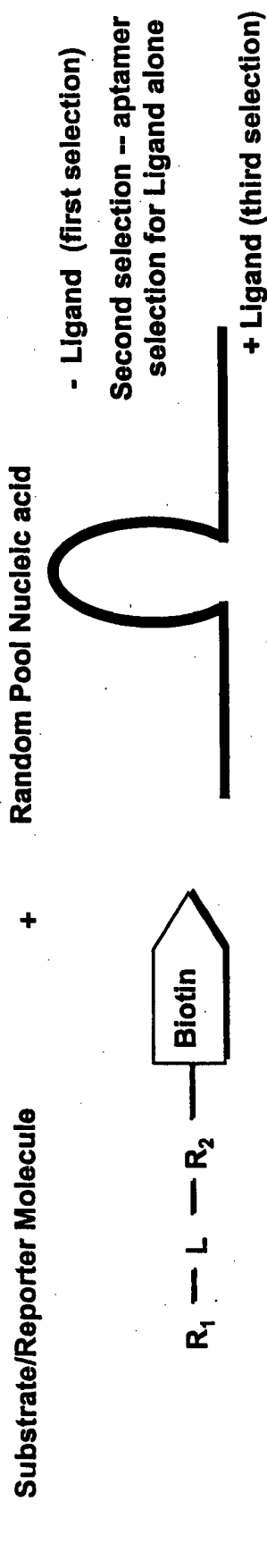
- Ligand (first round)
- + Ligand (second round)



Catalytic sequence is biotinylated

Reaction mixture passed over resin  
active molecules are trapped

**Figure 17c: Auto-ligation Nucleic Acid Sensor Molecules -  
Ligand dependent**



React substrate  
and random pool

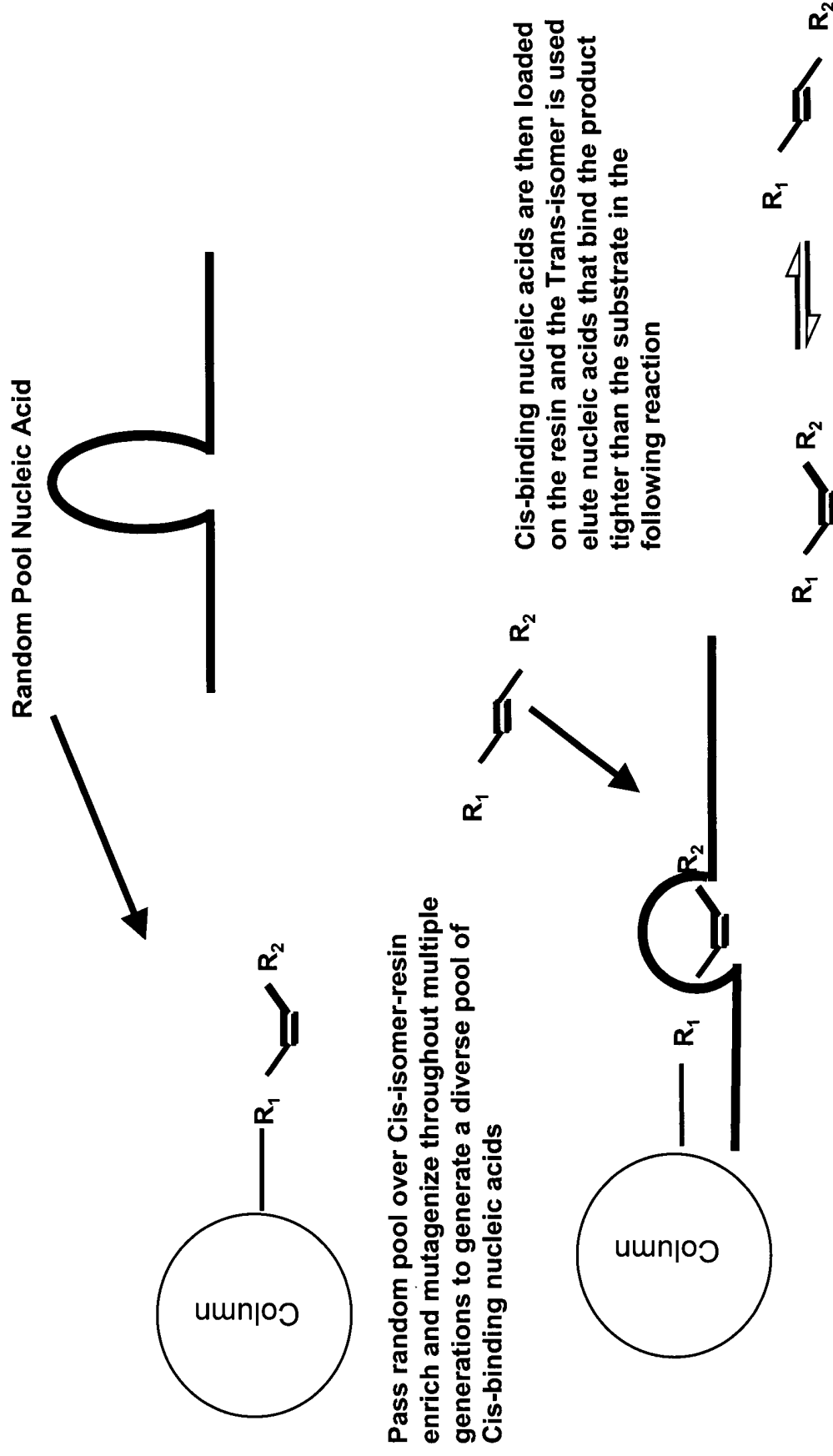
Scheme II

- Perform an entire selection as shown in first slide (in the absence of the Ligand)
- Mutagenize the winning pool
- Perform an entire selection using this pool with the requirement of Ligand binding.
- Mutagenize this pool
- Repeat original selection (for activity) in the presence of Ligand - countselect for molecules that react in the absence of ligand

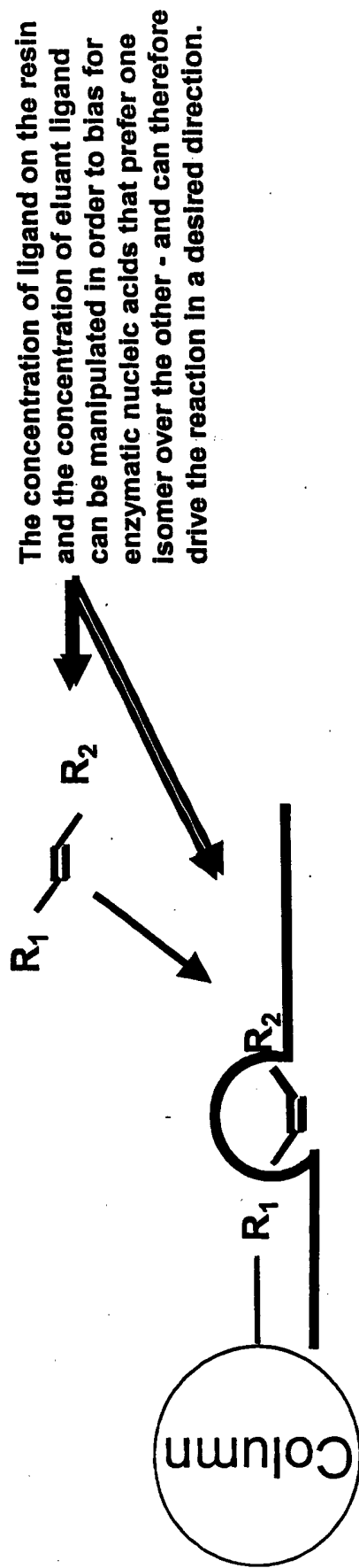
Catalytic sequence is biotinylated

Reaction mixture passed over resin  
active molecules are trapped

**Figure 18a: Isomerase Nucleic Acid Sensor Molecule – Selection Scheme**



## Figure 18b: Isomerase Nucleic Acid Sensor Molecule - Selection Scheme

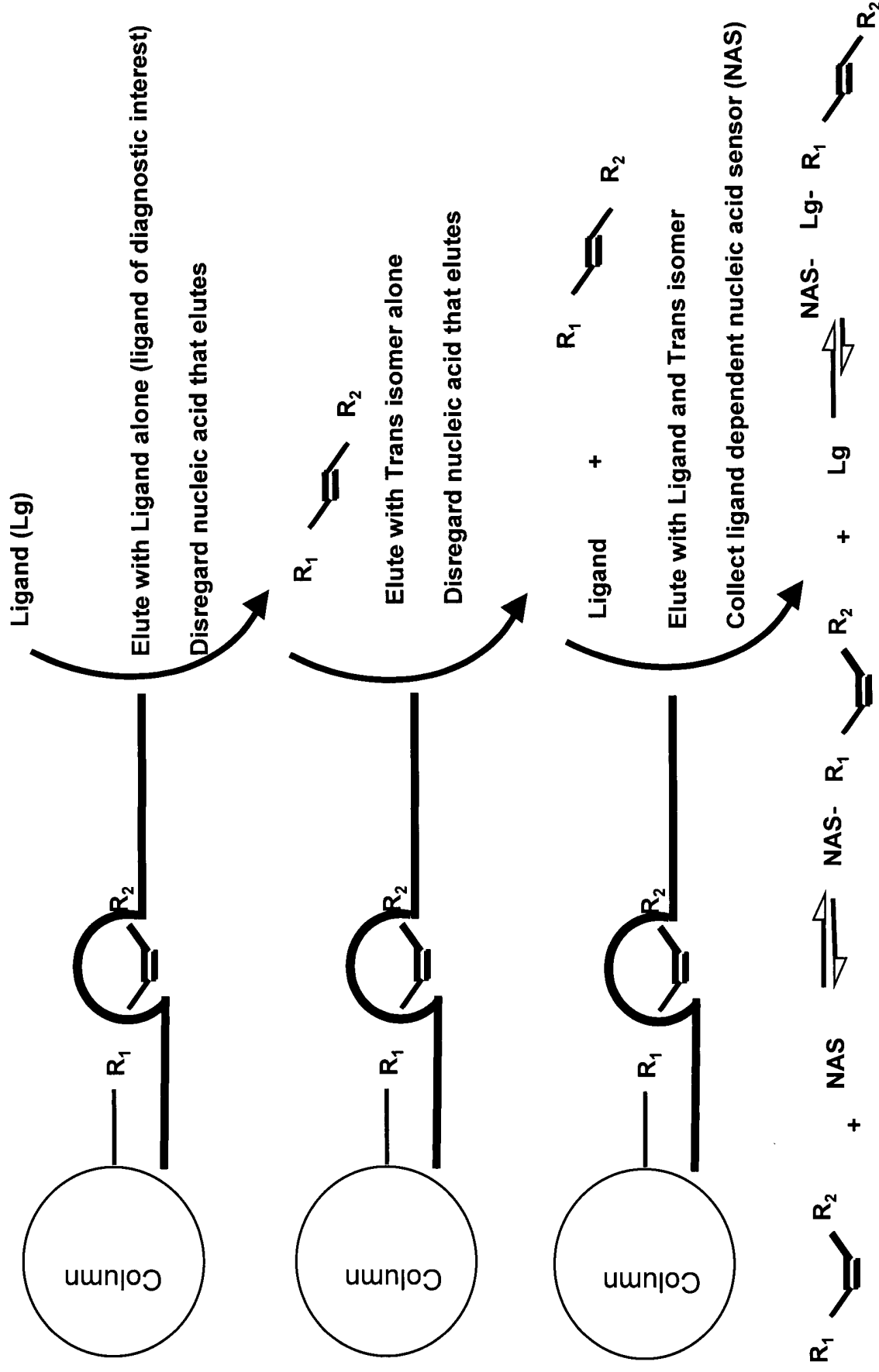


E.g. Selection for Cis-Isomer at 100  $\mu\text{M}$  - yield  $^{\text{cis}}K_d = 100 \mu\text{M}$   
 Elute with Trans-isomer at 0.1  $\mu\text{M}$  - yield  $^{\text{trans}}K_d = 0.1 \mu\text{M}$

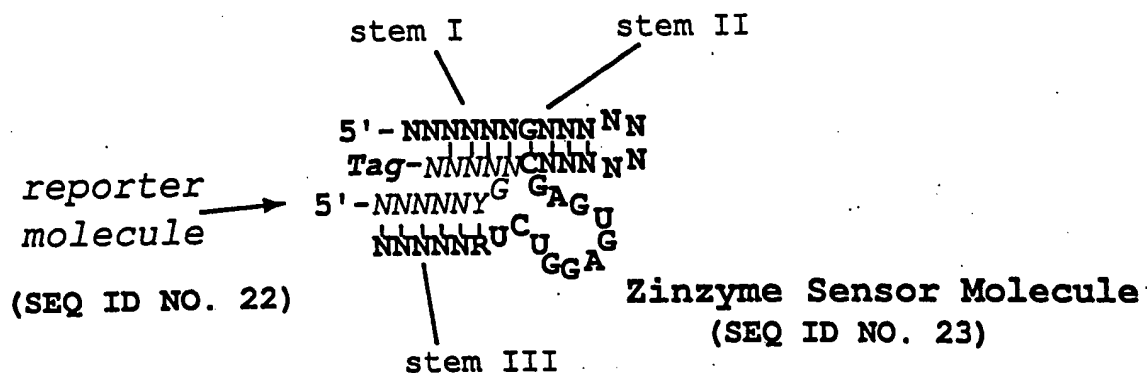
Isolate catalysts for the reaction below



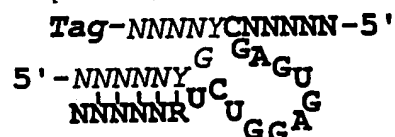
**Figure 18c: Isomerase Nucleic Acid Sensor Molecule - Ligand dependent**



# Zinzyme Sensor Molecule for detection of Nucleic Acid



Inactive Zinzyme sensor/  
reporter molecule complex  
(SEQ ID NO. 22)



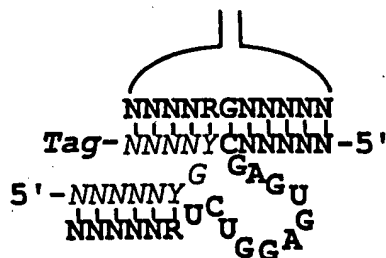
(SEQ ID NO. 24)

Target Signaling  
Molecule



Target Signaling  
Molecule

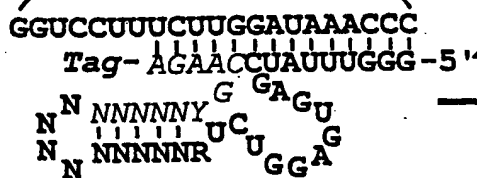
(SEQ ID NO. 25)



Active Zinzyme sensor/  
reporter molecule complex

Stem-loop III of HCV (SEQ ID NO. 26)  
(SEQ ID NO. 22)

Active HCV Zinzyme sensor/  
reporter molecule complex  
(SEQ ID NO 27)

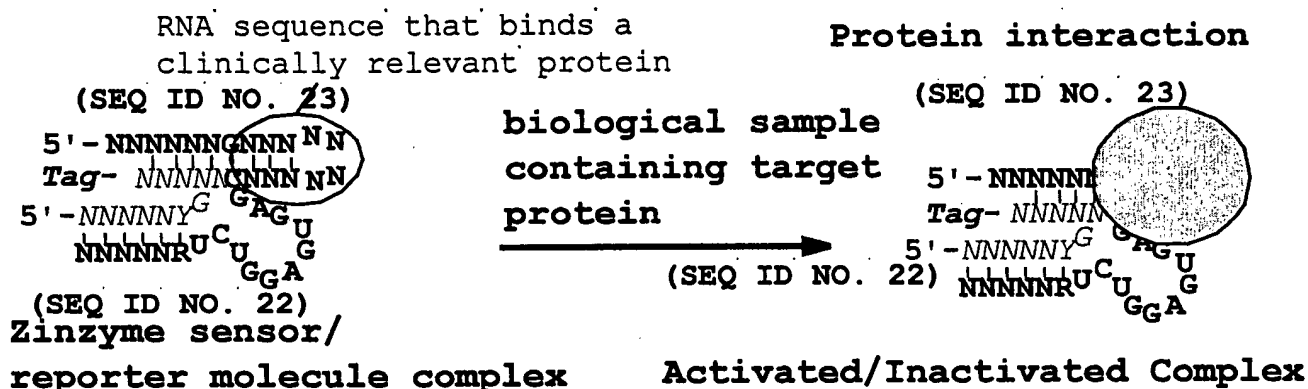
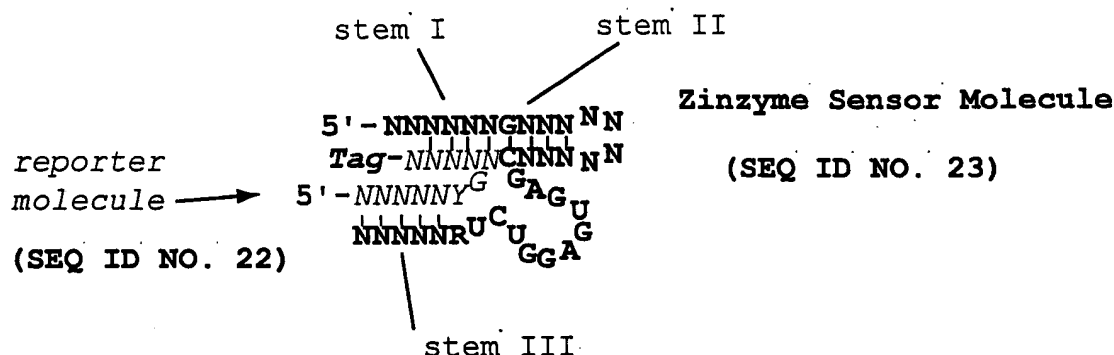


catalysis results in  
release of Tag-AGAAC  
for detection

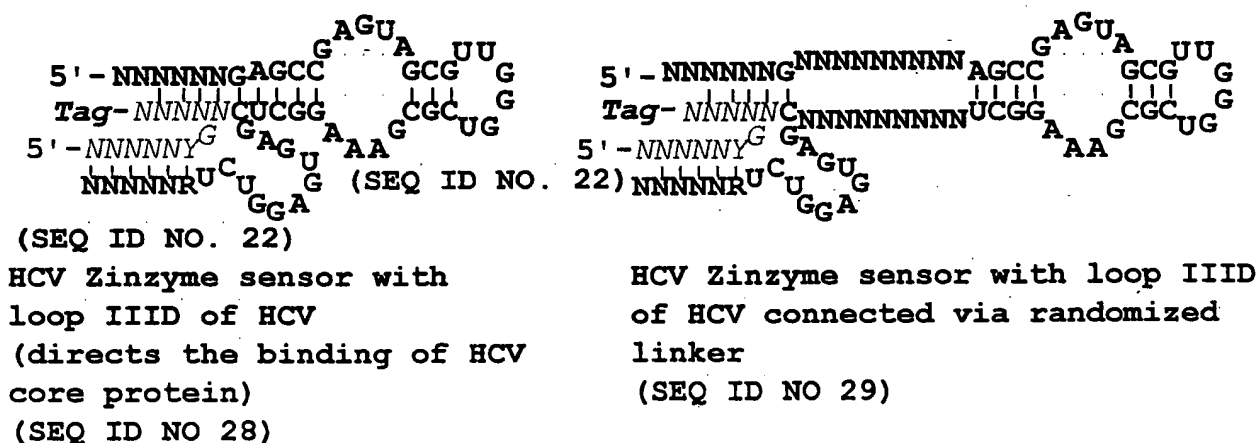
Zinzyme sensor can be attached to solid support/surface,  
for example at the 5'-end

FIG. 19

# Figure 20: Zinzyme Sensor Molecule for detection of Protein



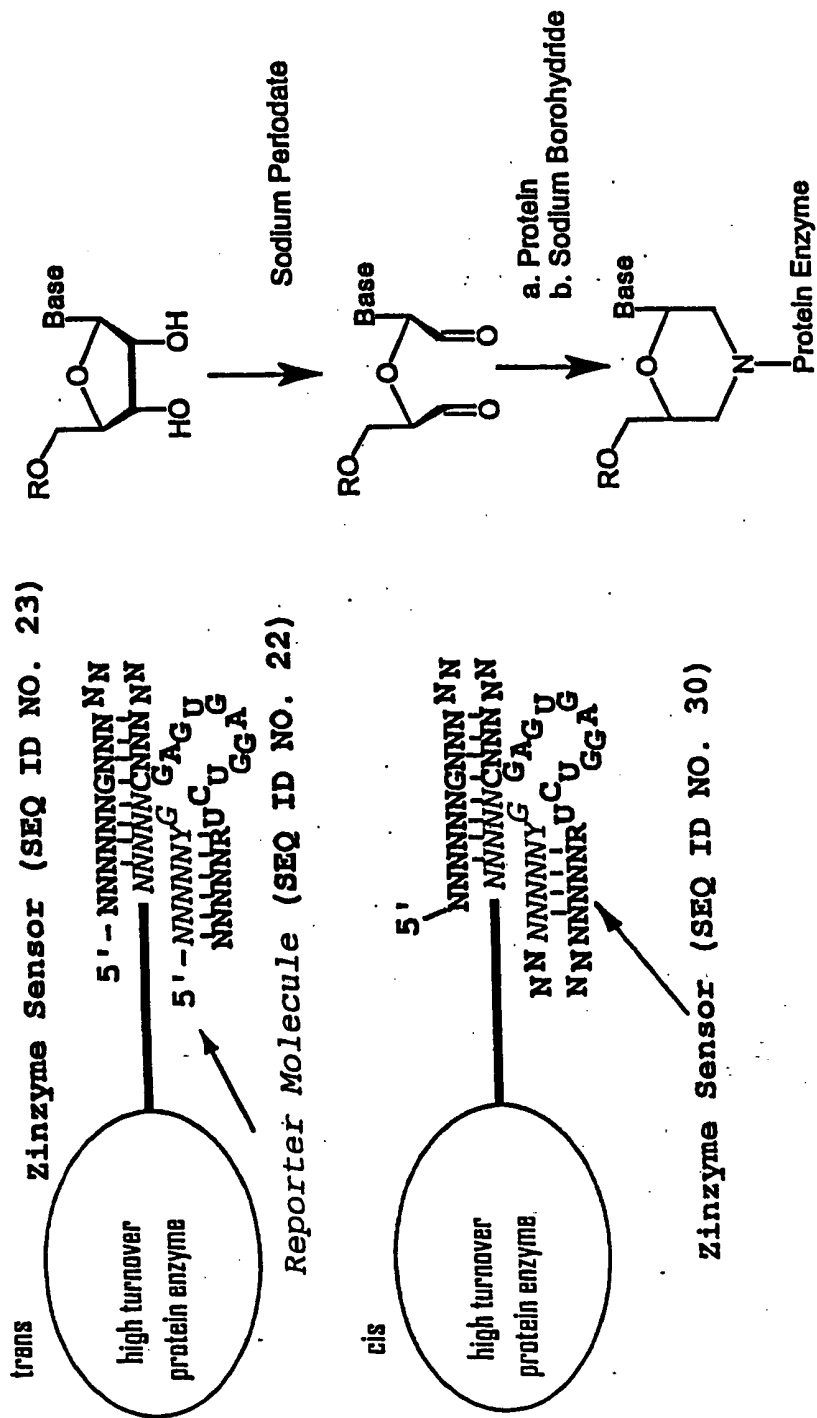
## Sensor/reporter complex for detection of HCV core protein



09877526 100001



# Zinzyme Sensor Molecule with protein enzyme reporter



R is oligonucleotide.

Protein can be attached via amino linker.

Alternately, R is phosphoramidite moiety for incorporation at 5'-end of oligonucleotide.

High turnover protein enzyme is, for example, Luciferase, Horseradish peroxidase, beta-galactosidase, alkaline phosphatase.

FIG. 21

# Amplification of signal via use of protein enzyme conjugate

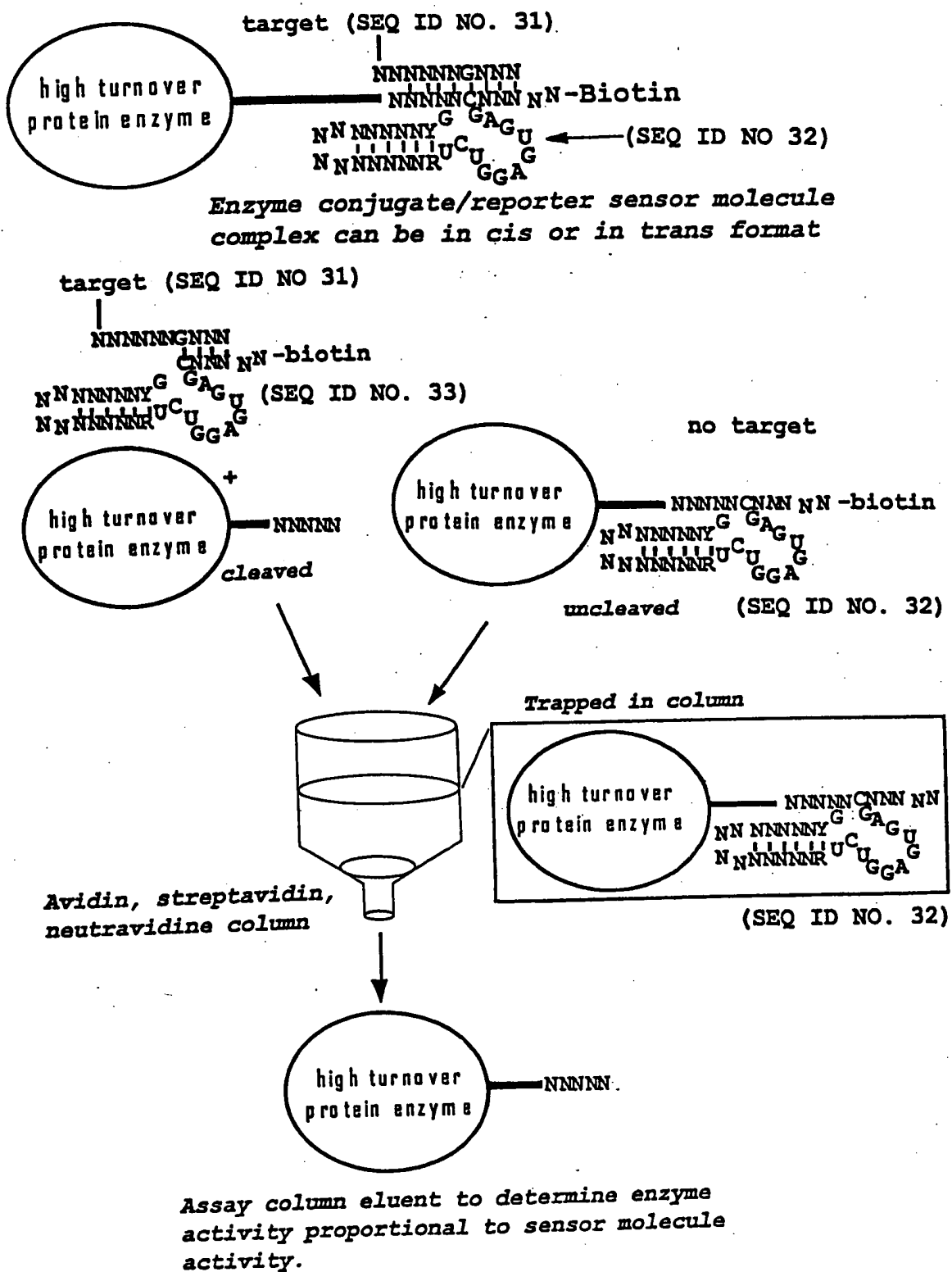
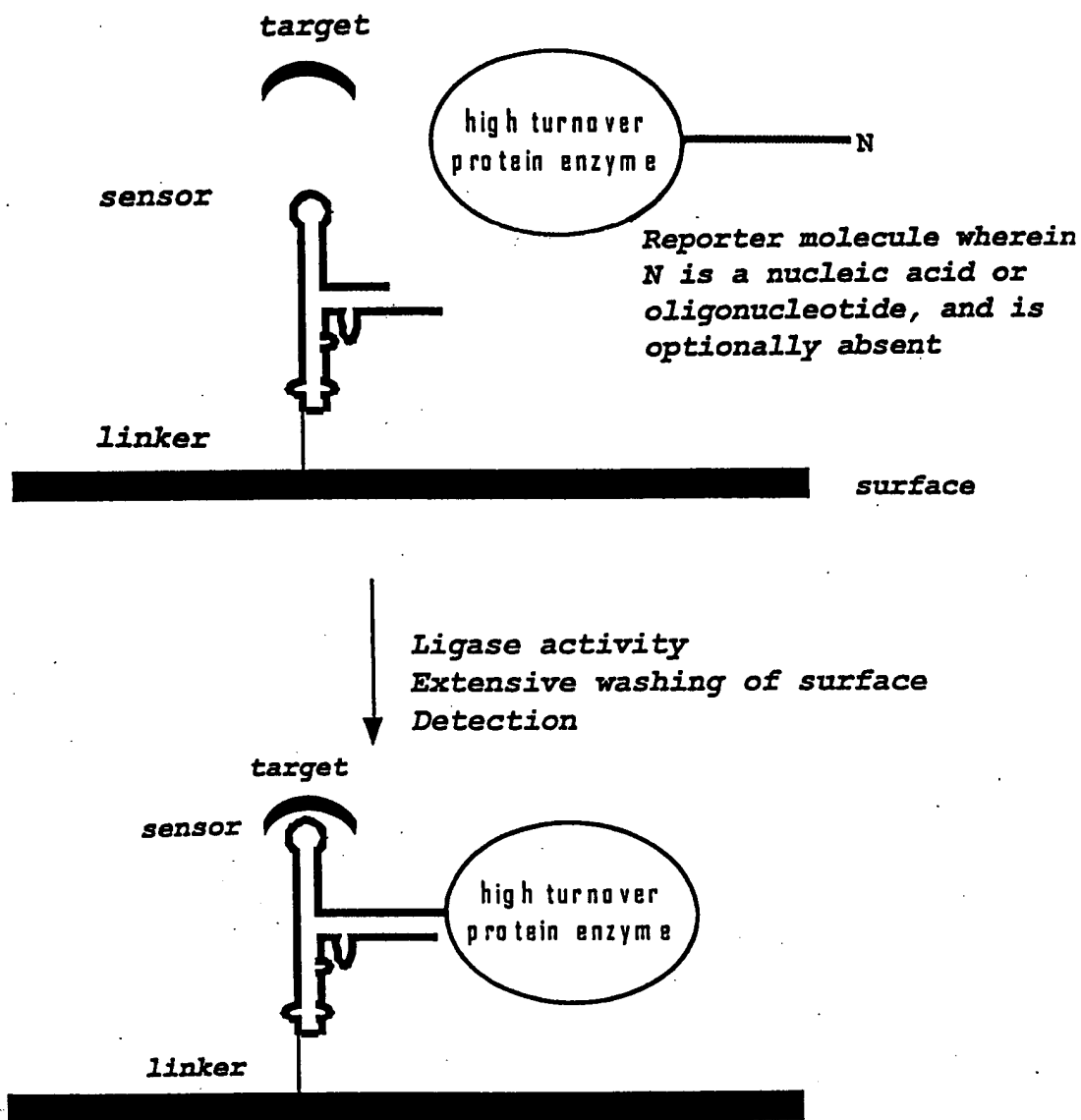


FIG. 22

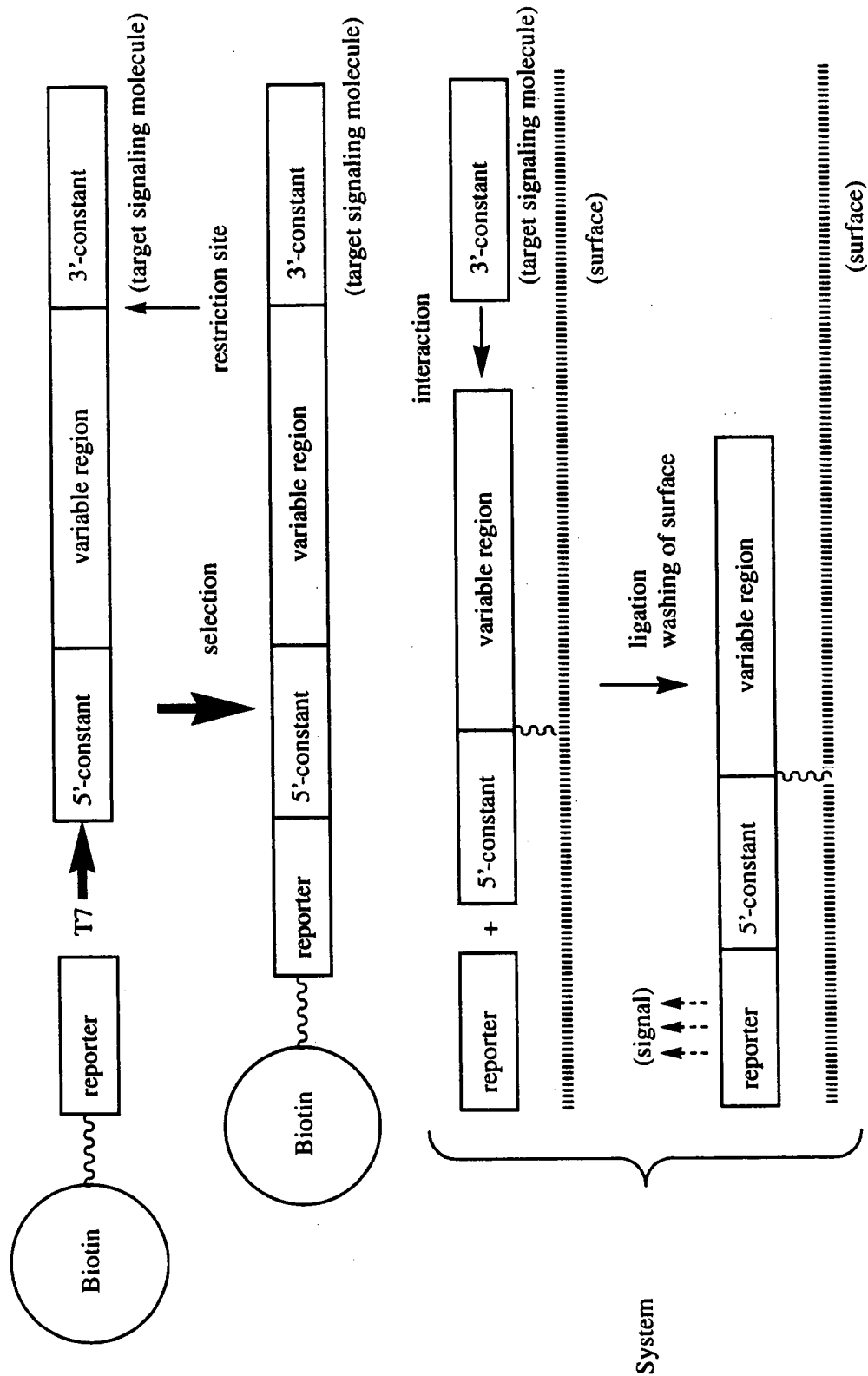
# *Ligase Sensor Molecule with enzymatic reporter*



Alternatively, a fluorescent or chemiluminescent based reporter molecule is used.

FIG. 23

Figure 24: Selection of Nucleic Acid Sensor Molecules with Ligase Activity



**Figure 25: Nucleic Acid Sensor Molecule-Based Electric Circuit**

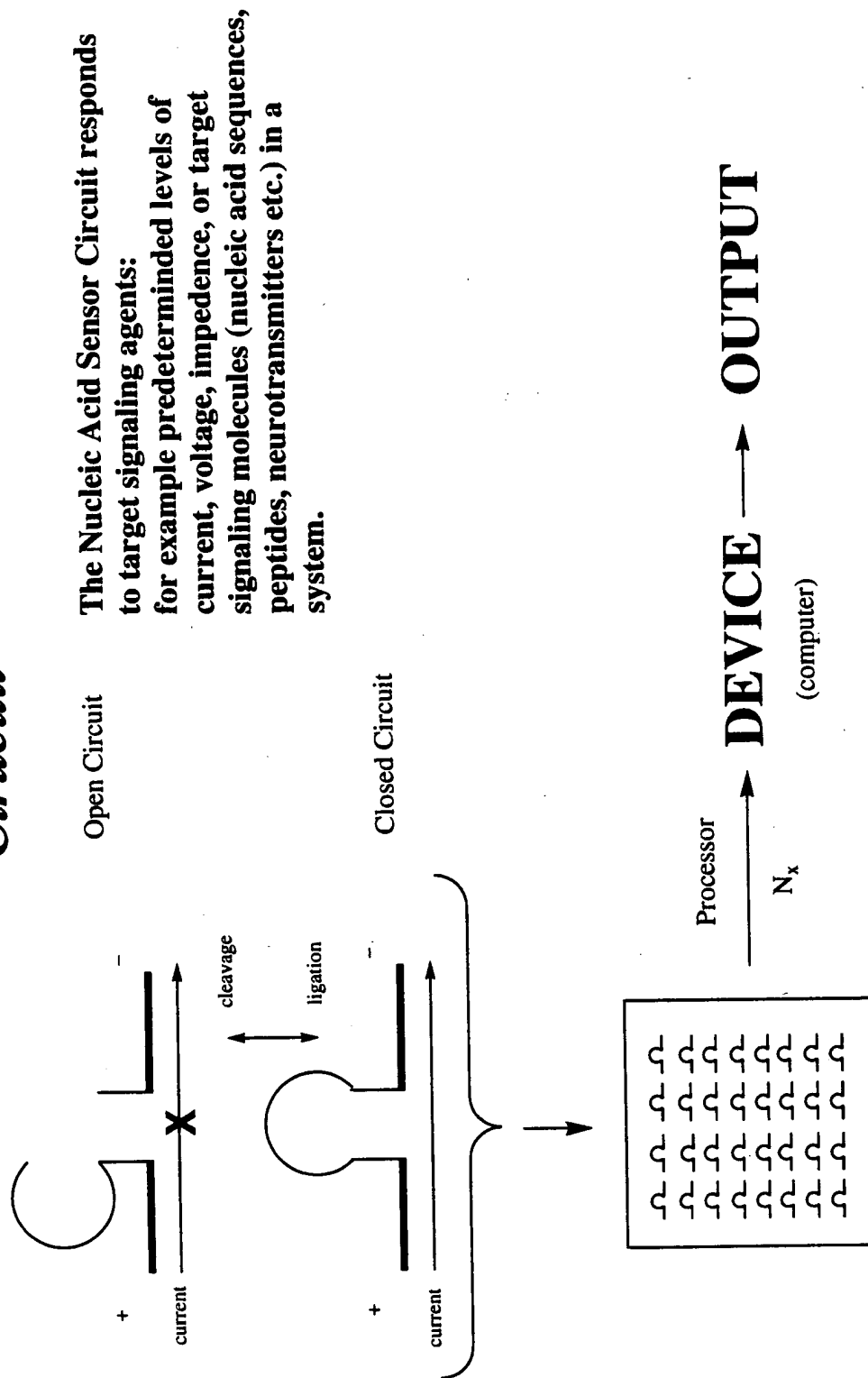
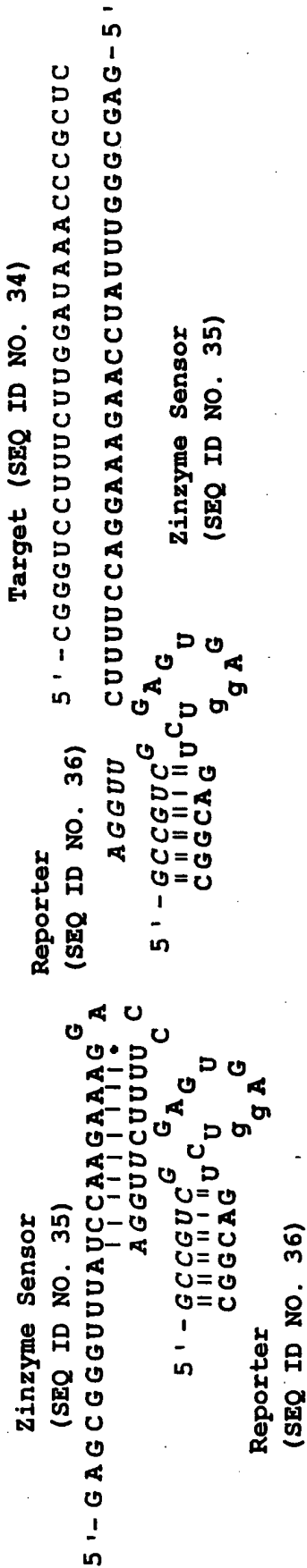
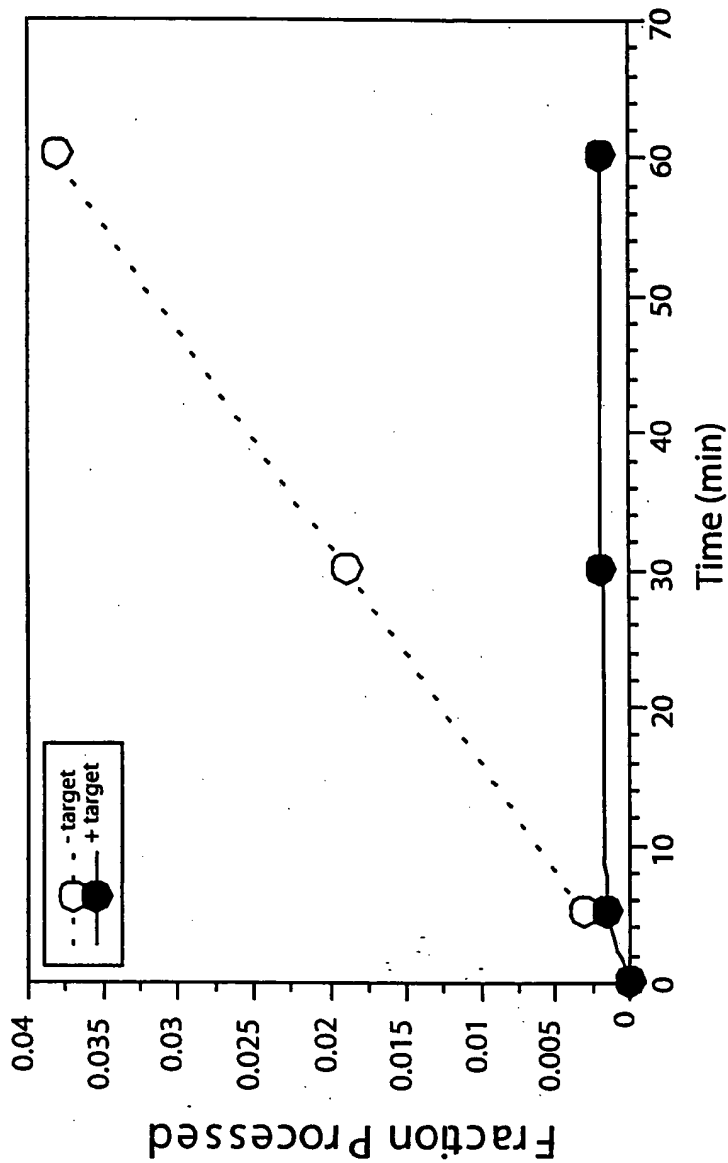


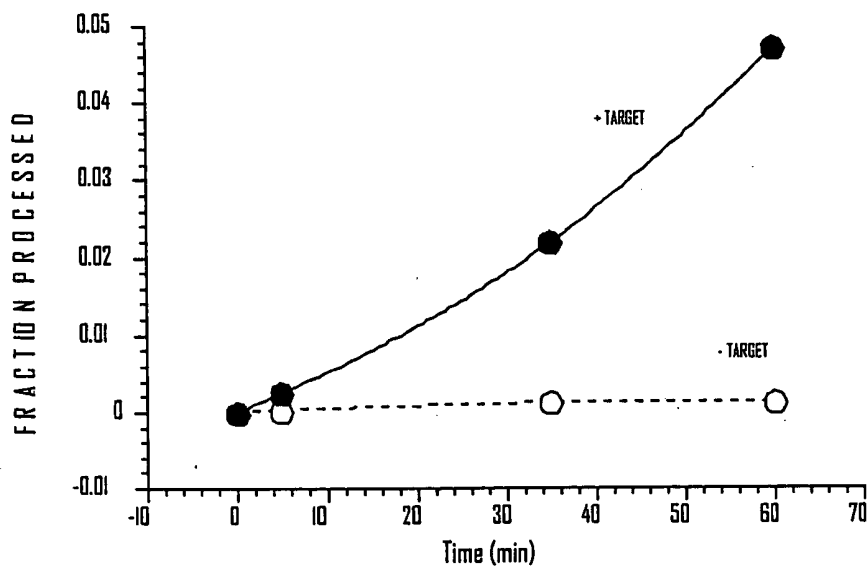
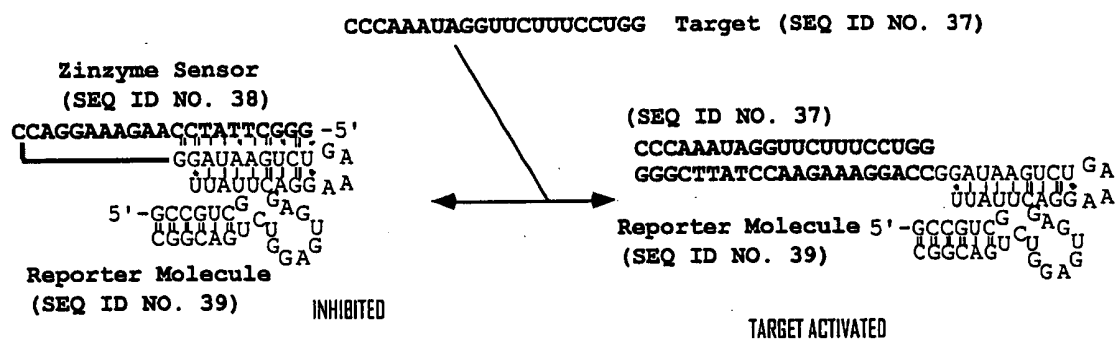
Figure 26: Target Inactivation of Zinzyme Sensor Molecule



ACTIVE  $\longleftrightarrow$  TARGET INACTIVATED



**Figure 27: Target Activation of Zinzyme Sensor Molecule**



0987556-10001

Figure 28: Erk modulated Nucleic Acid Sensor Molecule

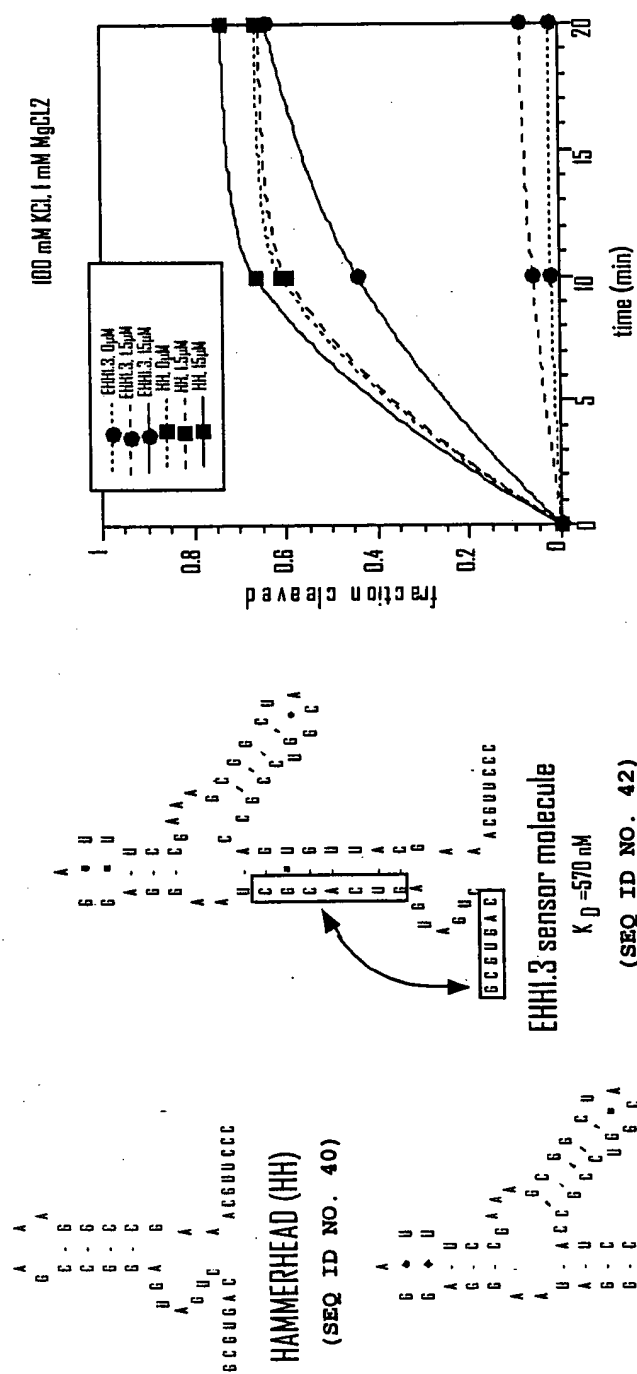






Figure 30

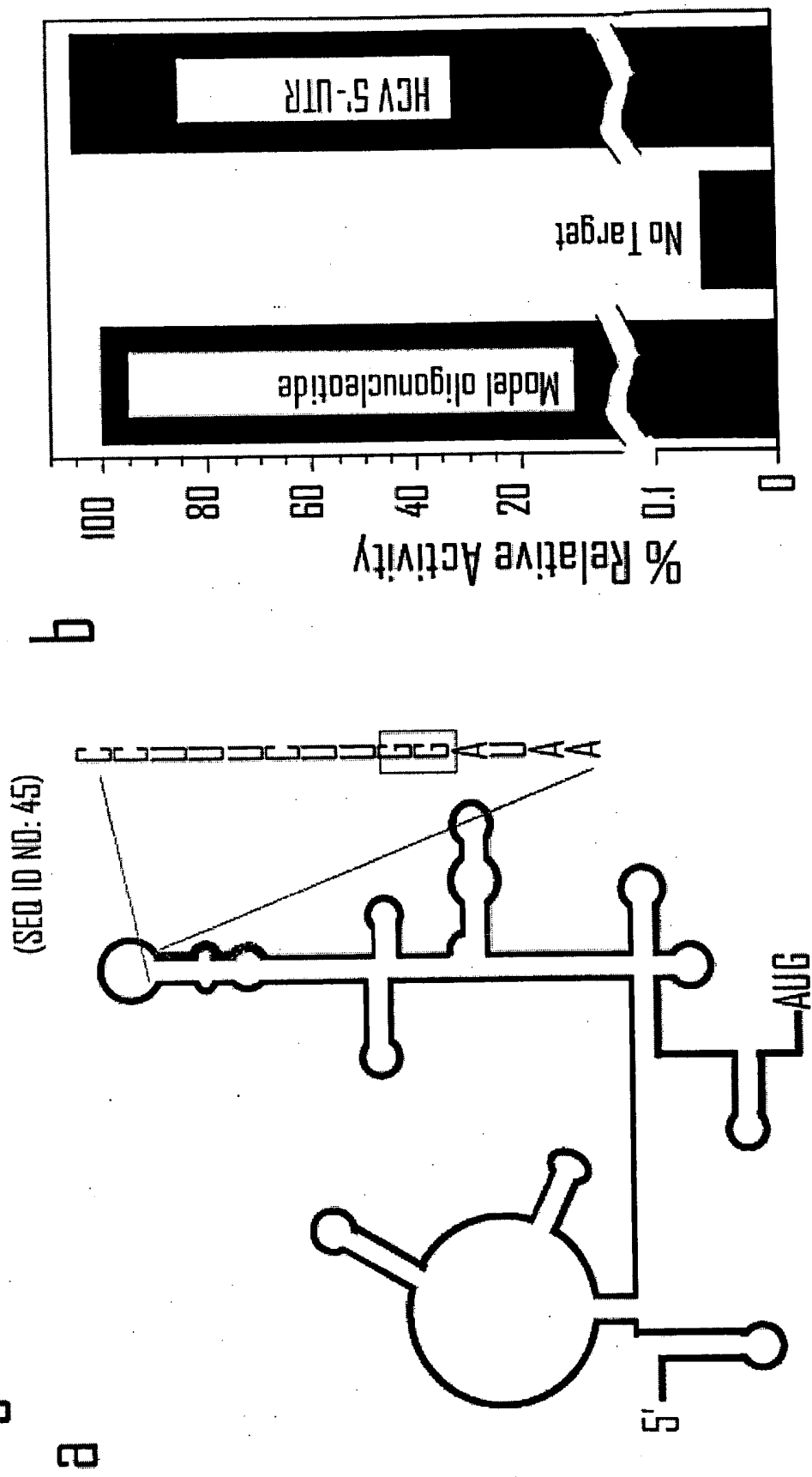


Figure 31

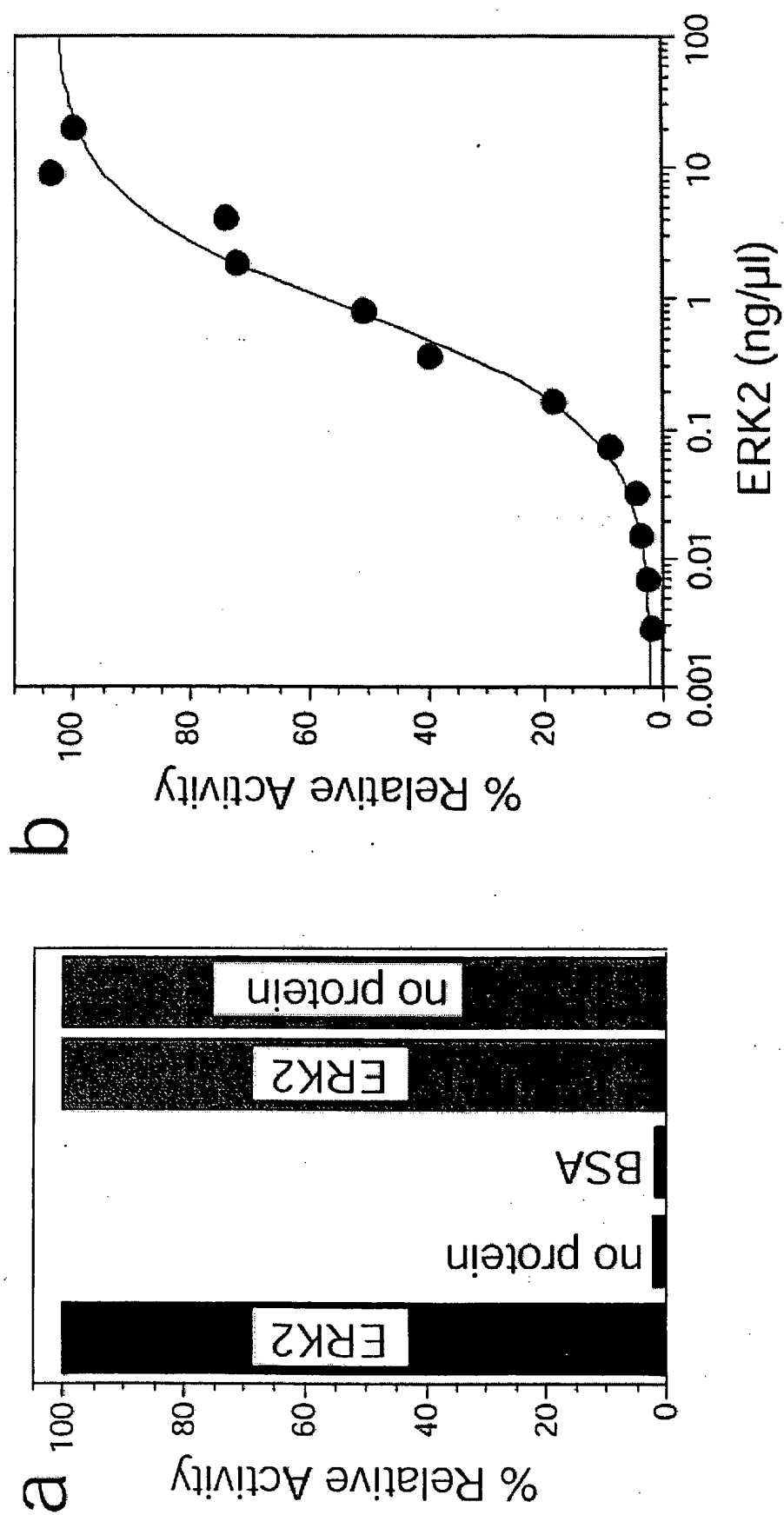
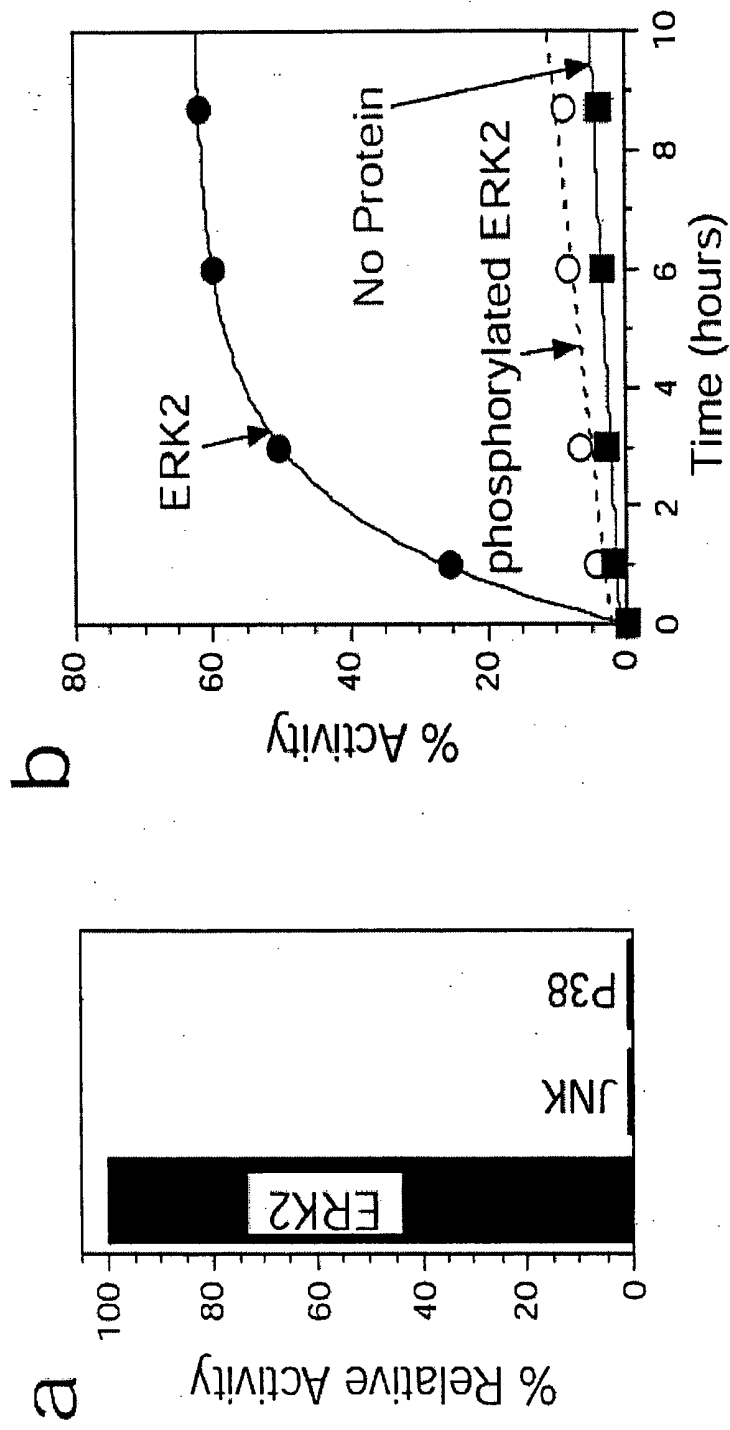
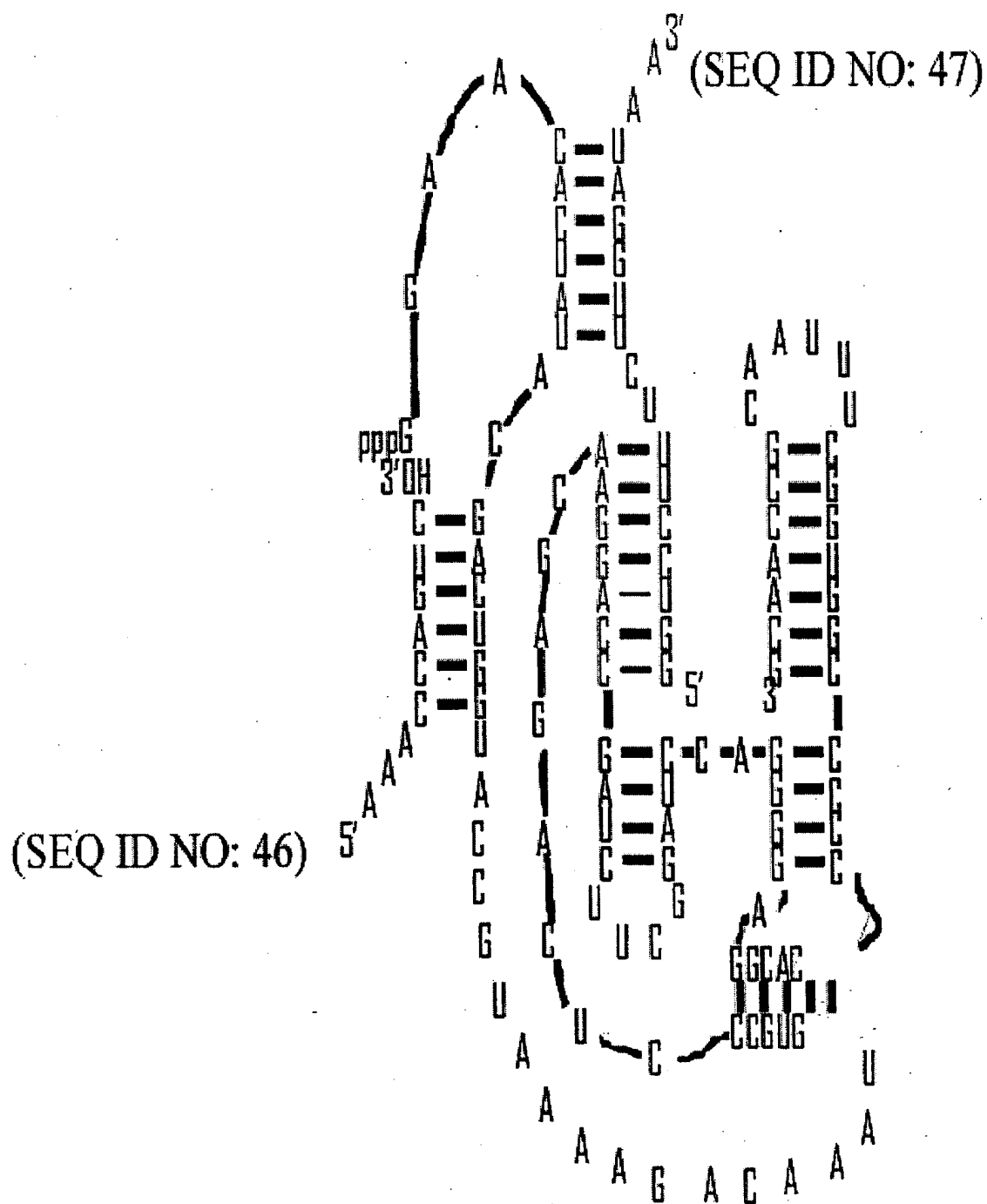


Figure 32



*Figure 33: Halfzyme Ligase*



0987526-100201

Figure 34: Secondary structure of HCV 5'-UTR

(SEQ ID NO 48)

5' GCGA GACACUCCACCAUAGAUCAUCC ACCCCCCUCCGGG GCG

